



AMERICAN UNIVERSITY OF BEIRUT

PARTICLE BOUND POLYCYCLIC AROMATIC  
HYDROCARBONS: SEASONAL VARIATION, SOURCE  
APPORTIONMENT AND CANCER RISK ESTIMATION

by  
FATIMA HUSSEIN HUSSEIN

A thesis  
submitted in partial fulfillment of the requirements  
for the degree of Master of Science  
to the Department of Chemistry  
of the Faculty of Arts and Sciences  
at the American University of Beirut

Beirut, Lebanon  
April 2018

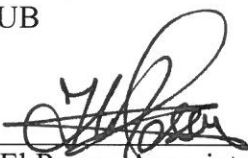
AMERICAN UNIVERSITY OF BEIRUT

PARTICLE BOUND POLYCYCLIC AROMATIC  
HYDROCARBONS: SEASONAL VARIATION, SOURCE  
APPORTIONMENT AND CANCER RISK ESTIMATION

by  
FATIMA HUSSEIN HUSSEIN

Approved by:

  
\_\_\_\_\_  
Dr. Najat A. Saliba, Professor  
Chemistry, AUB  
Advisor

  
\_\_\_\_\_  
Dr. Houssam El Rassy, Associate professor  
Chemistry, AUB  
Member of Committee

  
\_\_\_\_\_  
Dr. Issam Lakkis, Associate professor  
Mechanical Engineering, AUB  
Member of Committee

Date of thesis defense: April 2018

AMERICAN UNIVERSITY OF BEIRUT

THESIS, DISSERTATION, PROJECT RELEASE FORM

Student Name:

Hussein Hussein Fatima  
Middle Last First

Master's Thesis  
Dissertation

Master's Project

Doctoral

I authorize the American University of Beirut to: (a) reproduce hard or electronic copies of my thesis, dissertation, or project; (b) include such copies in the archives and digital repositories of the University; and (c) make freely available such copies to third parties for research or educational purposes.

I authorize the American University of Beirut, to: (a) reproduce hard or electronic copies of it; (b) include such copies in the archives and digital repositories of the University; and (c) make freely available such copies to third parties for research or educational purposes

after : **One ---- year from the date of submission of my thesis, dissertation, or project.**

**Two ---- years from the date of submission of my thesis, dissertation, or project.**

**Three ---- years from the date of submission of my thesis, dissertation, or project.**

Hussein Hussein  
Hussein

Signature

Date

02-05-2018

## ACKNOWLEDGMENTS

The completion of this thesis at the American university of Beirut would not have been possible without the support and encouragement of several people. Hence, I would like to take this opportunity to show my gratitude to those who have assisted me in a myriad of ways.

I would first like to express my sincere gratitude to my supervisor Professor Najat A. Saliba. A more supportive and considerate supervisor I could not have asked for. During the past three years she provided me with insightful academic guidance, encouraged me on developing research projects, and taught me how to be a patient and responsible person. There were many times where I had reached the crossroads and each time Prof. Najat was there to steer me towards the right path. She was always open to sit and listen to my troubles and always made me feel as if my work mattered. Her willingness to offer me so much of her time and intellect is the major reason for the completion of this accomplishment. I thank her for helping me to be ready for the next step in my life. It was a golden opportunity to be part of her research group.

I would like to acknowledge Dr. Houssam El-Rassy and Dr. Issam Lakkis for serving on my committee and providing helpful suggestions/comments on my thesis.

I would like to thank Professor Rabih Sultan for providing me with academic guidance and advice throughout my AUB journey.

I would like to thank the members of the AAL team, especially Ms. Rima Baalbaki for helping me to start the research work from zero. I was so lucky having you all; Miss Rima Baalbaki, Miss Lamis Al Aaraj, Mrs Rachelle El-Hajje, Dr. Ahmad El-Hellani Miss Sara El-Soussy, Miss Christina Haddad, Miss Maiassa Char, Miss Salwa Hjeir.

I would like to thank Mrs. Samira Atoui who gave me from her time to review, proofread this work and suggest helpful comments on my thesis.

I would like to thank my friend and roommate Linda Bechnak who was beside me during my hard times and happy moments. Special thanks to my friends Mohamad Hadi and Mohammad Tohme of being part of my graduate life at AUB.

Finally, I would like to dedicate this work to my dear family. My father Hussein Hussein I have no words to express my heartfelt gratitude and sincere appreciation for your constant encouragement, inspiration and wholehearted support at all the stages of my life. I appreciate all your trust and faith in me which helped in reaching this level. My mom who always kept me in her prayers, all the words cannot express how much I am thankful to your efforts, tenderness and caring that helped me finish this work with good health.

## AN ABSTRACT OF THE THESIS OF

Fatima Hussein Hussein for Master of Science  
Major: Chemistry

Title: Particle bound polycyclic aromatic hydrocarbons: seasonal variation, source apportionment and cancer risk estimation

**Introduction: background and aims:** Polycyclic aromatic hydrocarbons (PAHs) are ubiquitous pollutants released into the atmosphere as a mixture of different congeners due to incomplete combustion and pyrolysis of organic matter. Many PAHs and their derivatives are strongly potent carcinogens or mutagens to humans; therefore, their quantification and source identification in the environment are continuing concerns. Some studies in Lebanon reported short-term PAH levels at different locations. However, no studies up to our knowledge, evaluated the annual average and the seasonal variation of the 16 PAHs defined by EPA as the most potent at an urban background site. Hence, this study establishes a baseline level of PAHs at an urban representative site in Lebanon, evaluates their seasonal variation, apportion their sources and ultimately estimates their lifetime cancer risk.

**Methods:** Every six days Particle bound PAHs (P-PAHs) were sampled using a high-volume sampler at a representative urban site like AUB. The desired analytes were then extracted using an optimized method and analyzed using gas chromatography-mass spectrometry (GC-MS)

**Results:** The results showed that PAH levels were highly variable between the seasons throughout the year of 2017. The Highest levels of total PAHs were detected during winter ( $13.85 \pm 2.32$ ) ng/m<sup>3</sup> while lowest levels were detected in summer ( $6.32 \pm 0.74$ ). Consistently, the group 1 carcinogen Benzo[a]pyrene (B[a]p) followed the same trend of seasonal variation with an annual mean ( $0.75 \pm 0.23$ ) ng/m<sup>3</sup> lower than the annual recommended level determined by the European Union (1ng/m<sup>3</sup>). Among the different congeners of PAHs, the five-member aromatic rings known as possible or probable carcinogens were the dominant PAH congeners in the urban site. Incremental lifetime cancer risk (ILCR) scenario estimation showed that out of 10<sup>6</sup> population, one individual at this urban site would develop cancer due to daily exposure to ambient PAHs levels. PAH congeners can be emitted by different sources. Among these, gasoline, diesel and incineration were found to be the three major sources of PAHs determined by source apportionment with a contribution of 55.5%, 31.9% and 12.6%, respectively.

**Conclusion:** In short, our study establishes a baseline level of P-PAHs with an annual average of  $10.4 \pm 1.3 \text{ ng/m}^3$ . This level and the seasonal averages can be used as a benchmark to assess the contribution of additional sources in the different urban areas. The results also show a strong seasonal variation in P-PAH levels which indicates the effect of meteorological conditions and source emissions on their ambient concentrations. Positive matrix factorization (PMF) proved to be successful into partitioning the contributing sources according to their PAH markers because of the high number of samples provided to the model (n=55). Among the different PAH sources identified by this study, gasoline was found to be the major contributing source. Incremental life time cancer risk (ILCR) estimation for PAH-induced cancer exceeded the commonly acceptable threshold of  $10^{-6}$  and shows that there is a chance for an adult individual to develop cancer due to PAHs daily exposure.

# CONTENTS

ACKNOWLEDGEMENTS.....	v
ABSTRACT.....	vi
ILLUSTRATIONS.....	xi
TABLES.....	xiii
ABBREVIATIONS.....	xiv
Chapter	
I. INTRODUCTION.....	1
A. Particulate Matter.....	1
B. Polycyclic Aromatic Hydrocarbons(PAHs).....	4
C. Sources of PAHs.....	5
D. Physical properties of PAHs.....	5
E. Tropospheric transformation and degradation of PAHs.....	6
F. PAHs associated with particles in the atmosphere.....	9
G. PAHs toxicity.....	10
H. P-PAHs in Lebanon.....	13



I. Study objectives.....	16
J. sampling Site.....	17
<b>II. P-PAH SAMPLING, ANALYTICAL METHOD OPTIMIZATION AND VALIDATION.....</b>	<b>19</b>
A. Comparison between low-volume and high-volume sampling.....	19
B. Sample preparation of the sampled HVS filters.....	23
1. Materials.....	23
2. Filter extraction.....	23
3. Sample concentration.....	24
4. Solid phase extraction.....	25
5. Sample concentration.....	25
C. Instrumental analysis.....	26
D. Quantification.....	26
E. Method validation.....	28
1. Linearity.....	28
2. Limit of detection.....	29
3. Limit of quantification.....	30
4. Repeatability.....	30
5. Recovery.....	30
6. Blank repeatability.....	31
<b>III- P-PAHs AT AUB: RESULTS AND DISCUSSION.....</b>	<b>32</b>

A. Results: Variation of $\Sigma$ P-PAHs and P-B[a]p and individual P-PAHs.....	32
B. Discussion of Results.....	44
C. Classification of P-PAHs at AUB.....	46
D. Bap toxicity equivalent concentration.....	48
E. Incremental life time cancer risk .....	51
IV- SOURCE APPORTIONMENT OF P-PAHs USING POSITIVE MATRIX	
FACTORIZATION (PMF) AT AUB.....	53
A. Introduction to receptor modeling.....	53
B. Theoretical overview of Positive matrix factorization.....	54
C. PMF model optimization and validation.....	55
1. Signal to noise ratio.....	56
2. Observed/ predicted scatter plots.....	57
3. Residual Analysis.....	57
4. Bootstrap mapping.....	57
D. PMF output results.....	59
E. Discussion.....	61
V. CONCLUSIONS.....	64
VI. BIBLIOGRAPHY.....	66

## ILLUSTRATIONS

FIGURE		PAGE
I-1.	The different sizes of particulate matter along with and their different penetration pathways into the respiratory system.	3
I-2.	Pyrolysis followed by Pyrosynthesis of naphthalene starting with ethane	4
I-3.	Photolysis of tropospheric ozone to produce hydroxyl radicals	7
I-4.	Mechanisms of decomposition and transformation of Naphthalene in the troposphere during day time	8
I-5.	Formation of nitro-naphthalene during night time	9
I-6.	The 16 PAHs identified by the environmental protection agency (EPA)	11
I-7.	Metabolic oxidation of B[a]P followed by formation of B[a]P-DNA adduct leading to DNA mutation and inducing carcinogenesis	13
I-8.	Location of the sampling site	18
II-9.	The high-volume sampler, the PM <sub>10</sub> sampler impactor, and the HVS compartments constituting it	21
II-10.	The low volume sampler with a PM <sub>10</sub> impactor, and the setup	22
II-11.	Comparison of PPAH concentrations between high and low volume sampling	23
II-12.	Examples of the elution peaks for fluoranthene and pyrene of same molecular weight	27
II-13.	Example of elution peak of benzo(a)pyrene the group one carcinogen among the PAH family	28
II-14.	Examples of direct calibration curves of Acenaphthene and phenanthrene	29
III-15.	Temporal variation of the sum of P-PAH concentration on the sampled days	33
III-16.	Temporal variation of B[a]P (Group 1 carcinogen) concentration on the sampled days	34
III-17.	Seasonal variation for the average concentration of the 16 PAHs during the winter season	36
III-18.	Seasonal variation for the average concentration of the 16 PAHs during the spring season	37
III-19.	Seasonal variation for the average concentration of the 16 PAHs during the summer season	37

III-20.	Seasonal variation for the average concentration of the 16 PAHs during the fall season	38
III-21.	Correlation between sum of P-PAHs concentration and wind speed during the cold season	39
III-22.	Whisker and box plot of $\Sigma$ P-PAH during the twelve months campaign	40
III-23.	Whisker and box plot of $\Sigma$ P-PAH during the four seasons	41
III-24.	Whisker and box plot of P-B[a]p during the twelve months campaign	42
III-25.	Whisker and box plot of P-B[a]p during the four seasons	43
III-26.	Correlation between sum of P-PAHs and temperature during the cold and warm seasons	46
III-27.	Contribution of the three, four, five and six member rings to the total P-PAH sum	48
III-28.	Seasonal variation of B[a]p equivalent toxicity concentration	50
III-29	Percent contribution of the most carcinogenic PAHs to the B[a]P toxicity equivalence	51
IV-30	The three factor profiles for factors 1,2 and 3 determined by PMF and the fraction of the species in each	59
IV-31	Fingerprints (PAH markers) of the three factor profiles according to PMF results	60
IV-32	The portion contribution of the three factors to the total ambient P-PAH	61

## TABLES

Table	Page
I-1.	ATSDR / US-EPA priority PAHs, their phase distribution at ambient temperature, IARC Classification and TEF values
	12

## ABBREVIATIONS

ATSDR	Agency for Toxic Substances and Diseases Registry
AUB	American university of Beirut
AUH	American university hospital
B[a]p	Benzo(a)pyrene
CMB	Chemical mass balance
CO	Carbon monoxide
DNA	Deoxyribonucleic acid
EPA	Environmental protection agency
EU	European union
EPAQS	Expert Panel on Air Quality Standards
EHSRM	Environment and health safety risk management
EC	Elemental carbon
GC-MS	Gas chromatography-mass spectrometry
HPLC	High performance liquid chromatography
OH	Hydroxyl radical
IARC	International agency for research on cancer
IS	Internal standard
LOD	Limit of detection
LOQ	Limit of quantification
NAAQS	National ambient air quality standard
NO <sub>3</sub>	Nitrate radical
O <sub>3</sub>	Ozone

PAHs	Polycyclic aromatic hydrocarbons
PCA	Principle component analysis
P-PAHs	Particle bound polycyclic aromatic hydrocarbons
PM	Particulate matter
PM <sub>10-2.5</sub>	Coarse particulate matter
PM <sub>2.5</sub>	Fine particulate matter
PM <sub>0.1</sub>	Ultrafine particulate matter
PUF	Polyurethane foam
PMF	Positive matrix factorization
QC	Quality control
QA	Quality assurance
R <sup>2</sup>	Correlation coefficient
RSD	Relative standard deviation
RNA	Ribonucleic acid
S/N	Signal to noise ratio
SPE	Solid phase extraction
SOA	Secondary organic aerosol
TEF	Toxicity equivalence factor
UV	Ultraviolet
V-PAHs	Vapor phase polycyclic aromatic hydrocarbons
WHO	World health organization

## Chapter I

# POLYCYCLIC AROMATIC HYDROCARBONS IN AIRBORNE PARTICULATE MATTER

### **A-Particulate Matter**

Up to date, an extensive body of scientific evidence has shown that air pollution is linked to several ailments, due to the presence of toxic pollutants. In fact, exposure to these pollutants for either short or long term causes premature death and harmful effects on the human body, most of the time leading to many fatal diseases including cancer[1]. Even more, it is expected that by 2050, exposure to outdoor air pollution will take a lead among the environmental factors that cause premature mortality, thus surpassing malaria and water quality[2]. Recently, the World Health Organization (WHO) reported that in 2012, the indoor and outdoor air pollution led to 7 million premature deaths in the world, with 3.7 million deaths caused by outdoor air pollution[3]. Huge attention has been given to air pollution by scientists and great efforts have been made worldwide to reduce air pollution, especially after the establishment of Environmental Protection Agency (EPA) in 1970. Out of the many pollutants, EPA has defined National Ambient Air Quality Standards (NAAQS) for six principal pollutants-named "criteria" air pollutants. They are: Carbon Monoxide (CO), Sulfur Dioxide (SO<sub>2</sub>), Nitrogen Dioxide (NO<sub>2</sub>), Lead (Pb), ozone (O<sub>3</sub>), and Particulate Matter (PM)[4]. Among these "criteria pollutants", PM has received a high attention due to their strong association with mortality and morbidity cases from respiratory, cardiovascular , and cancer diseases [5, 6].

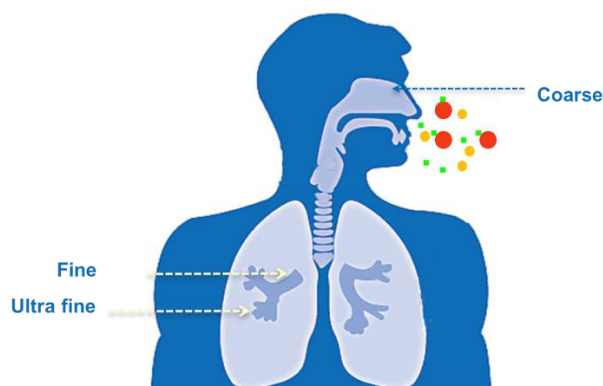
PM is a widespread air pollutant, consisting of a mixture of solid particles and liquid



droplets suspended in air. PM vary in sizes, they can range from few nanometers to several micrometers. Particles with diameters between 2.5 and 10  $\mu\text{m}$  ( $\text{PM}_{10-2.5}$ ) are known as coarse particles while those with a diameter between 2.5  $\mu\text{m}$  and 0.25  $\mu\text{m}$  ( $\text{PM}_{2.5-0.25}$ ) are known as fine particles. Particles that are less than 0.25  $\mu\text{m}$  in diameter are called ultrafine particulates (UFP)[7, 8]. Each PM mode has its own sources, chemical composition, respiratory penetration and atmospheric lifetimes. Coarse PMs mainly include crustal material, sea salts, and biological factors and are produced by mechanical processes unlike fine and ultrafine PMs which primarily contain elemental carbon, metals and organic compounds that are generated by combustion processes[8, 9].

The size and composition of these particles has been directly linked to being the main cause of health problems[10]. Generally speaking, the smaller a particle is, the deeper will penetrate to deposit on the respiratory tract at an increasing rate. To date, particles that have the most impact on human health have been acknowledged to those less than 2.5  $\mu\text{m}$  ( $\text{PM}_{2.5}$ ). These particles have longer lifetimes in air ranging from days to weeks compared to coarse particles ( $\text{PM}_{10}$ ) which have a lifetime of minutes to hours. Accordingly, the fine particles ( $\text{PM}_{2.5}$ ) can travel distances up to 1000 km higher by two orders of magnitude than coarse particles( $\text{PM}_{10}$ )[11]. While particles between approximately 5 and 10  $\mu\text{m}$  are most likely deposited in the tracheobronchial tree (upper respiratory track), those less than 2.5  $\mu\text{m}$  are deposited in the respiratory bronchioles and the alveoli (pulmonary interstitial sites) where gas exchange occurs[12, 13] (Figure.1). Eventually, those of the fine and quasi-ultrafine mode particles will escape into the blood stream and deliver harmful chemicals to the blood system to cause significant health problems including decreased lung function,

aggravated asthma, cardiovascular diseases, oxidative stress, and ultimately cancer [6, 14].

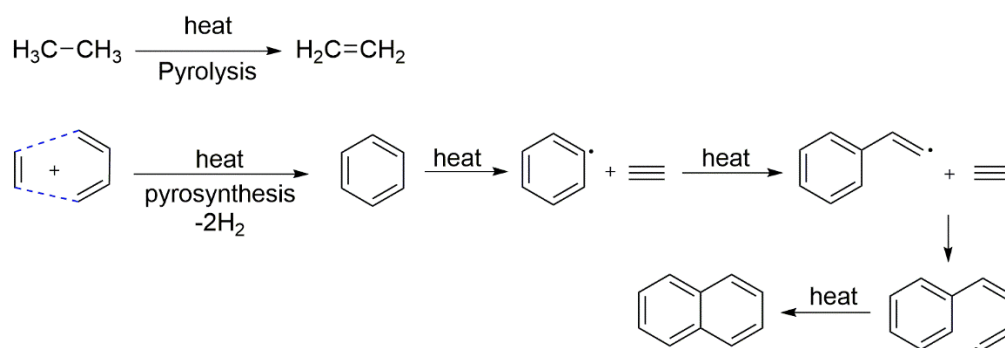


**Figure 1.** The different sizes of particulate matter and their different penetration pathways into the respiratory system.

Although the mechanism by which PM affects biological systems is still unclear, many studies has reported that metals act as possible mediators of PM induced airway injury and inflammation through the Fenton reaction[15-17]. Transition metals present in particles, especially iron, increase production of reactive oxygen species (ROS) in vivo[18]. As the release of ROS can result in cellular and tissue damage, it can thus initiate or exacerbate inflammation[19, 20]. However, a large fraction of ambient fine PM in many areas is derived from combustion processes and as such, contains significant amounts (up to 80–90% of UFP mass) of elemental carbon (EC) also called black carbon and organic carbon (OC)[21-24]. Part of the OC are polycyclic aromatic hydrocarbons (PAHs)[25]. PAHs are known to exert their toxicity after their biotransformation into toxic metabolites, which can be bound covalently to cellular macromolecules such as protein, Deoxyribo Nucleic Acid (DNA) and Ribonucleic Acid (RNA), causing cell damage, mutagenesis and carcinogenesis[26].

## B-Polycyclic aromatic hydrocarbons(PAHs)

PAHs are a class of semi volatile organic compounds, composed of at least two fused aromatic rings[27]. These compounds are widely distributed in the atmosphere and exhibit an increasing concern about their occurrence in the environment due to their carcinogenicity and mutagenicity[28]. Their ubiquitous presence in the atmosphere is mainly due to anthropogenic activities related to pyrolysis and incomplete combustion of organic matter such as paraffins, olefins, and aromatics. In general, during combustion or pyrolysis, where temperature exceeds 500°C, the organic compounds present in the fuel are cracked into smaller unstable free radicals, which in turn produce the first aromatic ring via several reaction mechanisms. Further reaction of this aromatic ring under high temperature and pressure conditions with small molecules such as acetylene (C<sub>2</sub>H<sub>2</sub>), ultimately leads to the growth of the aromatic system and the formation of larger multi-ring structures that are resistant to thermal degradation[Figure 2][29, 30].



**Figure 2.** Pyrolysis followed by Pyrosynthesis of naphthalene starting with ethane

## C-Sources of PAHs

In the ambient air of urban areas, PAH's occurrence is largely a result of anthropogenic emissions from many sources including fossil and biomass fuel burning power plants, road transport including diesel, and gasoline vehicles, waste incinerators, domestic heating, diesel generators, and industrial activities such as aluminum industry. The ambient existence of PAHs from biogenic sources including volcanic eruptions and forest fires is considered of negligible importance when compared to anthropogenic sources [29]. Different PAH emission sources usually produce different congeners. High levels of Fluorine, flouranthene, and pyrene along with moderate levels of high molecular weight PAHs benzo[b]fluoranthene and indeno[1,2,3-cd]pyrene mark the existence of oil combustion[29, 31, 32]. High factor loading of fluoranthene, benzo[ghi]perylene, benzo[a]anthracene, benzo[a]pyrene, and indeno[1,2,3-cd]pyrene is known to be attributed to gasoline emissions[29, 33-36]. In contrast to gasoline emissions, diesel emissions has a high factor loading of low molecular weight PAHs including flouranthene, phenanthrene, anthracene and pyrene [32, 37-43]. Power plant emissions are the major source of mainly heavy PAHs like benzo[k] fluoranthene, benzo[a]anthracene, benzo[a]pyrene and chrysene [40, 44].

#### **D-Physical properties of PAHs**

PAHs are semi-volatile organic compounds. The ring systems can be present in multiple configurations and several substitution structures[27] [Figure 3]. They have a relatively low solubility in water and are highly lipophilic. At ambient temperature, PAHs can be present in both particulate and vapor phases, depending upon their volatility. Compounds with 2-3 aromatic rings exist almost in the vapor phase, whereas compounds with 4 rings and above are particle-associated[27]. Atmospheric partitioning of PAHs is a form of particle/vapor

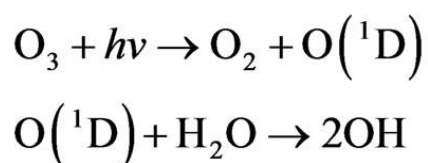
physical transformation that is affected by many atmospheric conditions among which meteorological parameters such as temperature and relative humidity are of high importance. On one hand, Yamasaki and coworkers[45] demonstrated the effect of temperature on the partitioning of PAHs using Fluoranthene, a 3-member aromatic ring PAH, as an example. They showed that lower temperatures cause a greater fraction of Fluoranthene to exist in the particle phase. This phenomenon can be explained by the enthalpy of adsorption on particles which is negative in this case. At low temperatures, the equilibrium of partitioning between the particle and the vapor phase shifts toward PAH adsorption on aerosols in the atmosphere. On another hand, Thibodeaux et al[46] investigated the effect of relative humidity on the partitioning of several PAHs. As the relative humidity is decreased down to 0%, sorption of PAHs increases. This increase is due to desorption of water where surface sites are opening up for sorption of the organic compounds. In short, temperature and relative humidity are two parameters with significant impact on the distribution of PAHs in the vapor and particulate phases.

### **E-Tropospheric transformation and degradation of PAHs**

PAH transformation and their fate in the environment is strongly influenced by whether the PAH is present in the vapor phase or is particle bound [47]. Vapor phase PAHs (V-PAHs) (2-3 aromatic rings) are highly susceptible to tropospheric photochemical reactions where they react with hydroxyl (OH) and nitrate (NO<sub>3</sub>) radicals leading to either their degradation or transformation[48]. In contrast, particle bound PAHs (P-PAHs) are less susceptible to tropospheric photochemical reactions due to the shielding effects of atmospheric organic compounds which can hinder their oxidation and promote their persistence in the troposphere[49, 50]. In fact, Atkinson, Arey, and their coworkers[51] investigated the

kinetics behind the chemical degradation of V-PAHs and P-PAHs. They reported that the reaction rate constants of V-PAHs by OH radicals are higher than those of P-PAHs by three orders of magnitude. For example, they have shown experimentally that tropospheric naphthalene, a compound highly present in the vapor phase, has a rate constant ( $K_{OH}$ ) of  $2.4 \times 10^{-11}$ , which is higher than that of Benzo(a)pyrene ( $K_{OH}$  of  $1.1 \times 10^{-14}$ ), a compound highly present in the particle phase. In short, since P-PAHs are less prone to degradation and are more persistent in ambient air than V-PAHs, special attention is given to P-PAHs in atmospheric PAH studies.

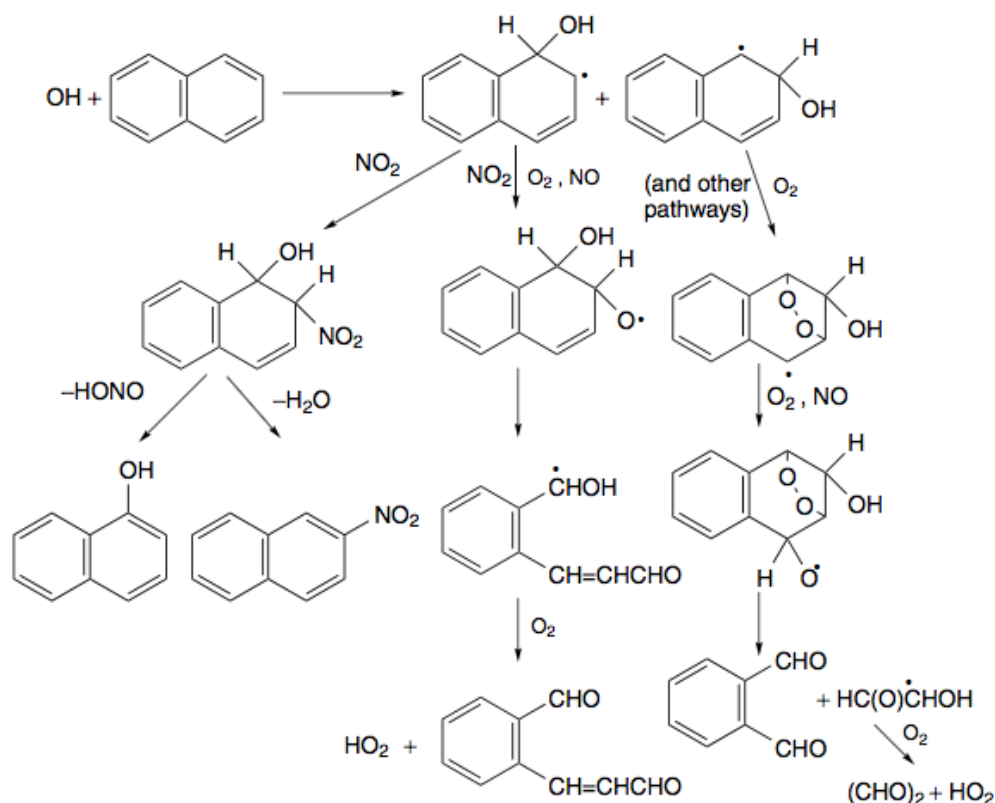
Among many factors that affect the fate of V-PAHs in the environment, atmospheric photochemistry appears to be one of the most important parameters in the degradation of V-PAHs. These atmospheric chemical reactions can be classified into two categories based upon the presence and absence of sunlight. During the day, OH radicals are dominantly present in the troposphere as a result of the increased rate of ozone ( $O_3$ )'s photolysis (Figure3)



**Figure 3.** Photolysis of tropospheric ozone to produce hydroxyl radicals[52]

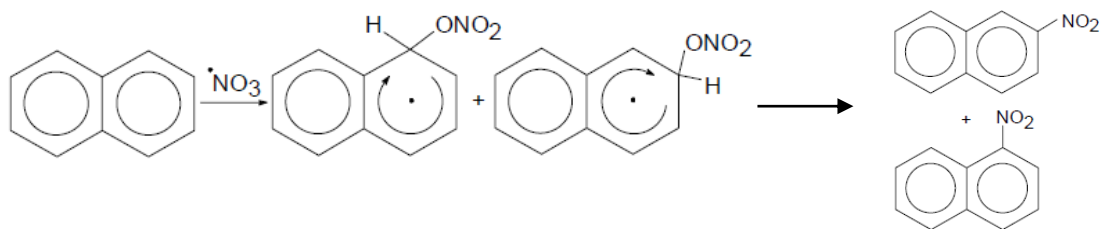
These tropospheric OH radicals interact with V-PAHs and trigger either their degradation or transformation into carboxylic acids, aldehydes, ketones, and quinones within a loss

lifetime of V-PAHs ranging between minutes to days. Above 40% of these products are degradation products (Figure 4).



**Figure 4.** Mechanisms of decomposition and transformation of Naphtalene in the troposphere during day time[53, 54]

In contrast, at night, as a result of the decreased photolysis, tropospheric  $O_3$  are highly available to interact with  $NO_2$  and form  $NO_3$  radicals. In turn,  $NO_3$  radicals interact with V-PAHs to highly produce transformation products mainly Nitro-PAH derivatives within a V-PAH loss lifetime similar to that seen during day time (Figure 5). It is important to note here that these transformation products are highly mutagenic as well as stable and persistent (lifetime = years) which may have strong impact on health and environment.[47]



**Figure 5.** Formation of nitro-naphthalene during night time[51]

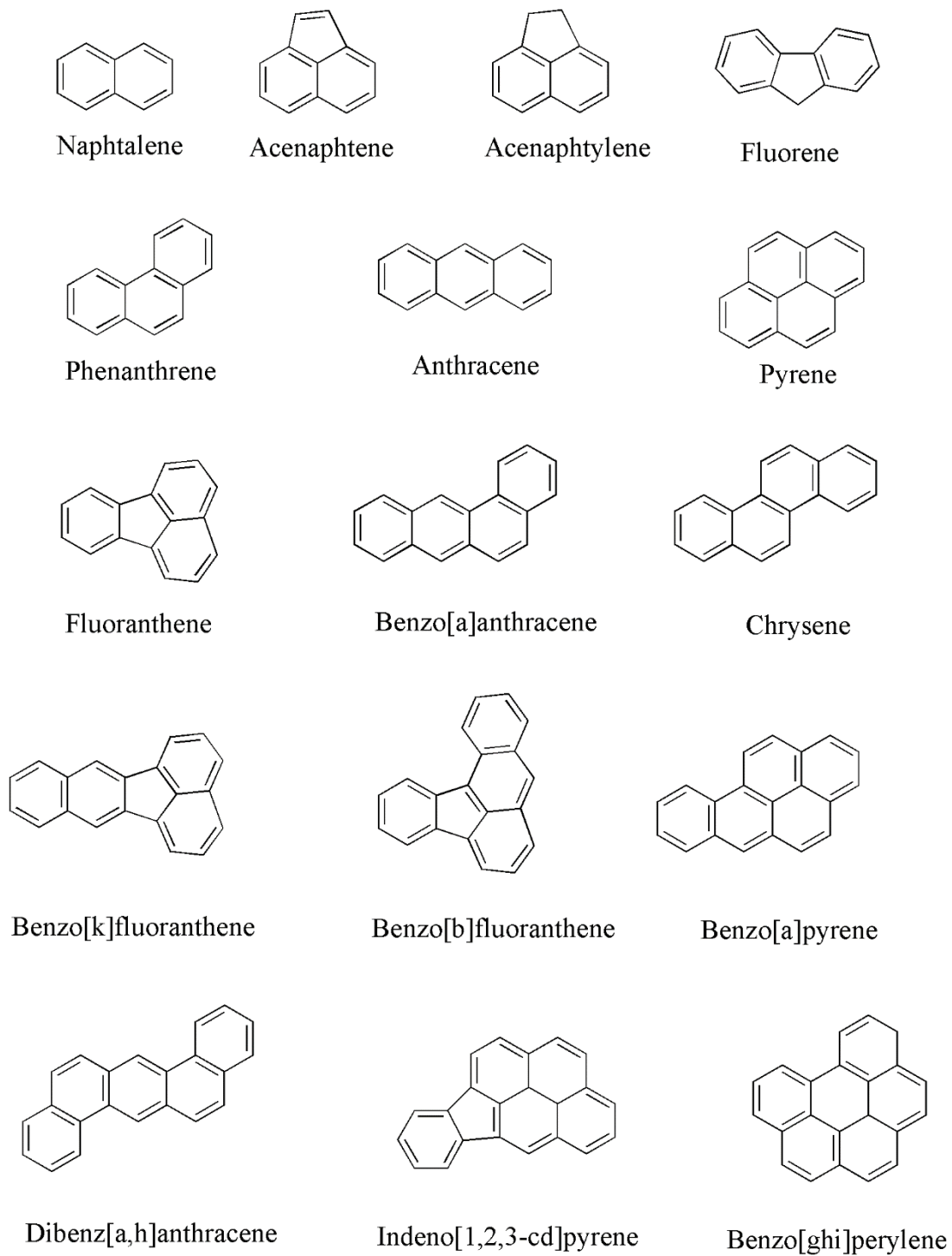
### **F-PAHs associated with particles in the atmosphere**

It has been shown that in ambient air semi volatile PAHs have a high affinity to small size particulate matter (PM<sub>2.5</sub>) known as fine PM. These fine PM include diesel exhaust particles or particles generated from incomplete combustion processes. Fine PMs are mainly composed of a major core of amorphous allotrope of carbon (EC) to which organic compounds, predominantly PAHs, are adhered[55-57]. Through their direct interaction with PAHs, these carbonaceous particles exhibit a potential inhibiting factor on the reactivity of PAHs. This inhibitory effect is due to the slow diffusion of tropospheric oxidants ( $\text{OH}$ ,  $\text{NO}_3$ ,  $\text{O}_3$ ) as well as to the slow diffusion of atmospheric oxidants into the bulk particle unlike the case of V-PAHs as previously mentioned. In addition, oxidation of volatile organic compounds in the atmosphere releases secondary organic aerosols (SOA) which in turn can shield fine PMs and trap the PAHs inside a highly viscous and hydrophobic medium. Once trapped, these PAHs can no longer evaporate and shield from atmospheric oxidation. This synergy between PAHs and SOA particles does indeed have important impact on increasing their residence time as in the atmosphere, thus leading to longer-range transport[58, 59]. However, direct photolysis of P-PAHs with ultraviolet (UV) sun light radiation is a potentially important process. The major degradation products due to their photolysis are epoxides and quinones[51].



## **G- PAHs Toxicity**

The PAH family includes more than 100 different compounds of various structures and toxicity. Accordingly, the assessment of toxicity for the numerous PAHs in the atmosphere is a complex process. In response, The US Environmental Protection Agency (EPA) has defined 16 unsubstituted PAHs as priority pollutants, based upon their toxicological profile as reported by the Agency for Toxic Substances and Diseases Registry (ATSDR) (Figure6)[60]. These 16 congeners were included in the priority list being suspected to be more harmful than others and exhibiting the highest concentrations in air of all the analyzed PAHs [29]. Among those congeners, benzo[a]pyrene (B[a]P), a five member aromatic ring almost found in the particulate phase, has been extensively studied and has been classified by the international Agency for research on cancer (IARC ) as a human carcinogen (Group 1)[61]. In response to its carcinogenic property, the European Union (EU) has set an air quality standard for B[a]P in the air of  $1 \text{ ng/m}^3$  [62]. However, in the United Kingdom B[a]P annual average concentration was recommended by the Expert Panel on Air Quality Standards (EPAQS) not to exceed  $0.25 \text{ ng/m}^3$  [63]. Moreover, other PAHs have been classified as probable human carcinogen (Group 2A) and possible human carcinogen (Group 2B) which are mostly found in the particulate phase [61, 64, 65]. Table 1 shows IARC classification for the 16 priority PAHs and their corresponding Toxic equivalency factor (TEF). TEF expresses the toxicity of individual chemicals within a family of compounds. A TEF of 1 is given to the most toxic chemical in the family (in this case B[a]p); less toxic chemicals are given TEFs of 0.1, 0.01 or 0.001 based on their relative toxicity to B[a]p (Table1).



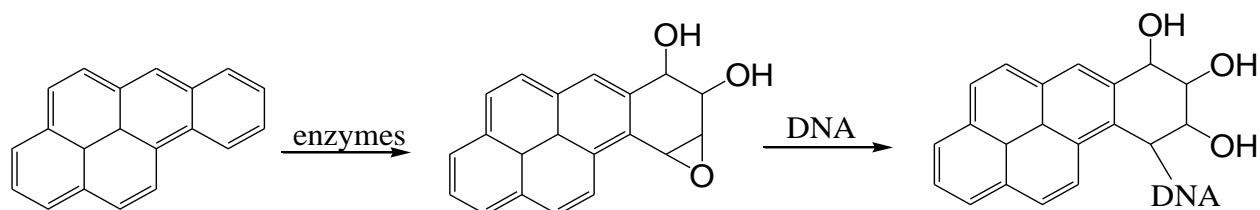
**Figure 6.** The 16 PAHs identified by the environmental protection agency (EPA)

**Table 1.** ATSDR / US-EPA priority PAHs, their phase distribution at ambient temperature, IARC Classification and TEF values

<b>PAH</b>	<b>Gas/Particle Distribution</b>	<b>IARC Classification</b>	<b>TEF</b>
<b>Naphthalene</b>	Gas phase	Group 2B	0.0001
<b>Acenaphthylene</b>	Gas phase		0.001
<b>Acenaphthene</b>	Gas phase	Group 3	0.001
<b>Flourene</b>	Gas phase	Group 3	0.001
<b>Phenanthrene</b>	Particle and Gas phase	Group 3	0.001
<b>Anthracene</b>	Particle and Gas phase	Group 3	0.01
<b>Flouranthene</b>	Particle and Gas phase	Group 3	0.001
<b>Pyrene</b>	Particle and Gas phase	Group 3	0.001
<b>Benzo[a]anthracene</b>	Particle phase	Group 2B	0.1
<b>Chrysene</b>	Particle phase	Group 2B	0.01
<b>Benzo[k]flouranthene</b>	Particle phase	Group 2B	0.1
<b>Benzo[a]pyrene</b>	Particle phase	Group 1	1
<b>Benzo[g,h,i]perylene</b>	Particle phase	Group 3	0.01
<b>Dibenzo[a,h]anthracene</b>	Particle phase	Group 2A	1
<b>Indeno[1,2,3-cd]pyrene</b>	Particle phase	Group 2B	0.1

PAHs can affect the human health through various toxicity mechanisms, including genotoxicity, immunotoxicity, developmental toxicity, and carcinogenesis. Long-term exposure to PAHs can induce epigenetic modifications. PAHs contribute to epigenetic remodeling through DNA methylation, histone modification, and microRNA silencing. Through series of reactions catalyzed enzymatically, reactive PAH metabolites attach to cellular proteins and DNA. The formed PAH-DNA adducts of different structural conformations can lead to induced mutations or inactivation of the tumor suppressor genes; which ultimately lead to DNA damage and carcinogenesis (Figure 7) [26, 66-68]. Studies

also show that PAHs are potent immunotoxic agents that can lead to immunosuppression through different pathways[69, 70]. Evidence is supporting that PAHs can cause developmental toxicity through prenatal and postnatal exposure to PAHs [71-73].



**Figure 7.** Metabolic oxidation of B[a]P followed by formation of B[a]P-DNA adduct leading to DNA mutation and inducing carcinogenesis[74]

### **H-Pollution in Lebanon: introducing the rationale of studying P-PAHs**

The Mediterranean region is known for high pollution episodes due to its enclosed geography. It is situated between the Saharan desert of Africa and the very heavily populated and highly industrialized European continent, which makes it prone to aerosol accumulation. In addition, its proximity to the three continents (Africa, Asia, and Europe) makes out of it an intersection of the air masses that meet in this region. Thus, this Mediterranean basin is characterized by a high humidity, long summers and stagnant winds that originate from eastern Europe. Furthermore, the intense solar radiation present in this area contributes to the high photochemical reactions that leads to pollutants' formation and/or transformation. These photochemical reactions and pollutants accumulation are much higher in the eastern side of the Mediterranean basin than the western side due to its dry summers and very low precipitation. Not only that, but also the eastern side is characterized by particulate matter dust outbreaks originating from Saharan and Arabian

desserts during the fall and spring seasons which thus makes it a controversial region at the pollution level[75-80]. Among the east Mediterranean cities, Beirut (7800 persons/km<sup>2</sup>), the capital of Lebanon, a city geographically located between the Mediterranean coast from the west and mount Lebanon from the east (35°28'790"E, 33°54'139"N) is the city where 88% of the 6 million population are present . In its densely populated city, the roads are often congested with high levels of traffic throughout the day, which can cause delays of up to 30 minutes at some intersections[81]. Moreover, construction operations are a normal part of city activity; with an urban population growth rate of 3.2%, as opposed to the global average of 2.1%[82]. New residence buildings are constructed to accommodate the increasing city population. Construction activities also occur for the maintenance of the country's outdated infrastructure[83]. Furthermore, due to inadequate power production capacity, the national electric company Electricité du Liban (EDL) has instigated a rotating power outage regime across the country[84]. In turn, citizens have resorted to privately-owned diesel generator to compensate for the needed electric power. It is worth noting, however, that Lebanon lacks heavy industrial facilities, with the exception of a number of thermal power plants and low-duty industrial factories situated away from the main cities of the country[85].

Over the past decade, several instantaneous and long-term studies were conducted in Beirut to investigate the levels, seasonal variation, and composition of atmospheric particulate matter and have reported that ambient PM levels in the greater Beirut area of Lebanon consistently exceed the WHO guidelines by up to 273% for PM10 and around 100% for PM2.5[75, 76, 78, 79, 86]. Furthermore, the daily PM10 and PM2.5 concentrations have

been associated with increased respiratory and cardiovascular emergency hospital admissions[87].

Moreover, Reactive oxygen species assay conducted in Beirut have shown that when compared gram by gram, the intrinsic toxicity of roadside PM<sub>2.5</sub> are relatively greater than the roadside PM<sub>2.5</sub> studied in Los Angeles[88]. All these studies show the levels of PMs however, the track of the ambient carcinogenic composition i.e P-PAHs over a significant period affected by various emissions is still unclear. In fact, Shihadeh et al. studied P-PAHs emissions from private diesel generators after a 3-hr electricity power outage in Beirut-Hamra between January 2010 and January 2012. They reported an increase over the local background level of approximately 40% of airborne P-PAH in the Hamra area of Beirut [89] . Another study, which was conducted by Daher et al. in 2012 over a period of two months in the summer, reported P-PAHs levels near a major road in beirut to be 11.5 ng/m<sup>3</sup> compared to levels below the detection limits at American university of Beirut (AUB) [86]. In this study, low volume sampling (LVS) technique was used for collection and analysis. Baalbaki et al. studied Gas and particle PAHs for winter and summer in 2015 at three different urban sites and reported that PAH values vary between 13.15 and 91.88 ng/m<sup>3</sup>[90]. In this study, the high-volume sampling (HVS) technique was used to collect air samples. Although these studies clearly show that PAH levels in different sites are ubiquitous, up to our knowledge, no baseline of PAH sources and concentrations has been determined yet. In fact, a baseline study is of prime importance. It provides a reference point to monitor and compare PAH levels released by local anthropogenic sources at different urban locations. To be used as a point of reference, it is necessary to respect two

criteria: conduct the study at an urban background site and over a long period of time. Unlike the cross-sectional study conducted by Daher et al who performed their study at a specific location near the freeway, Shihadeh et al. performed a longitudinal study at a specific location. Although these studies have succeeded in evaluating PAH levels released by certain specific sources in specific areas of Lebanon, they have not determined a P-PAH ambient concentration baseline affected by combined emission sources such as gasoline, diesel, heavy fuel oil and incineration at a representative site. In addition, there is a lack of a long-term study that investigates the impact of meteorological factors on P-PAHs' seasonal variation. Therefore, our study has shifted the attention towards defining such a baseline in Beirut. Once such a baseline is established, it is then important to explore the different sources of P-PAHs and to investigate their portion of contribution into the overall P-PAH level in the ambient air. Therefore, a receptor model known as positive matrix factorization (PMF) is used to further look into PAH source apportionment in the urban background site. In order for this model to be highly efficient, it is important to input a high number of samples; a criterion met by our study (n=55). In addition, in order to assess the efficiency of different sampling and analysis techniques used by the aforementioned studies, our study evaluates the efficiency in collecting and quantifying PAHs using LVS and HVS as methods of collection and establishes an optimized analytical extraction method.

### **I-Study objectives**

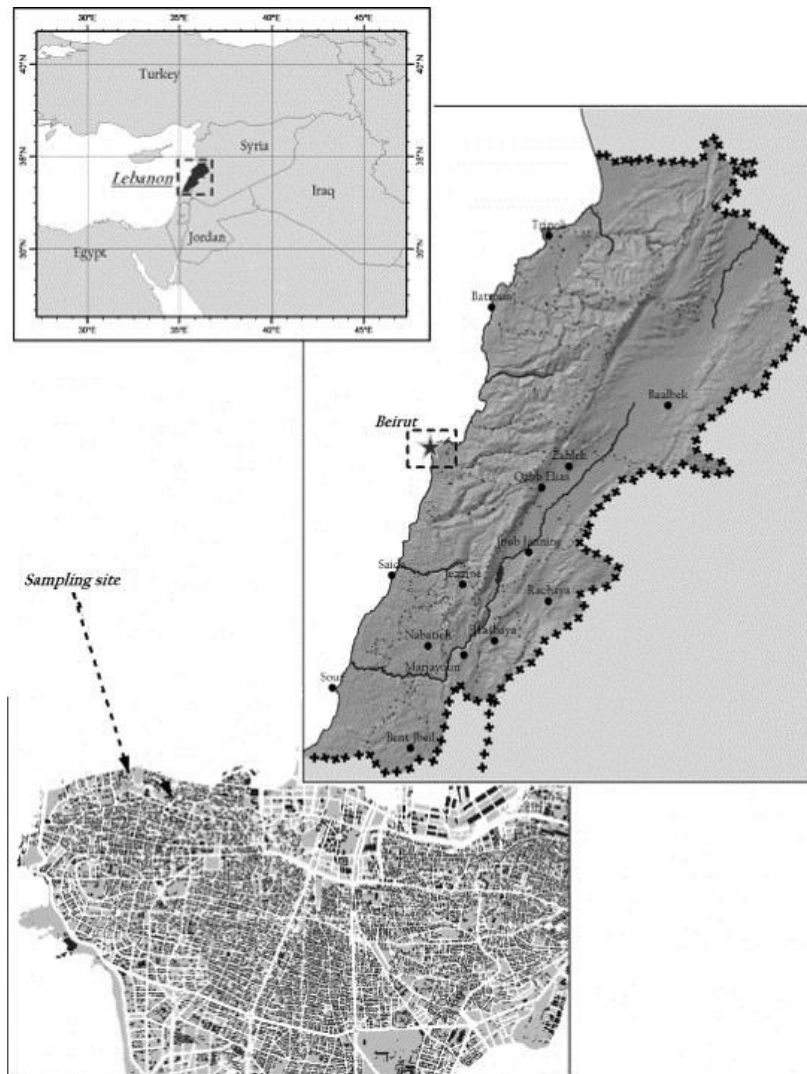
To establish a baseline level of P-PAHs at an urban representative site, determine P-PAHs seasonal and intermonth variation over a period of a year, explore the main factors affecting

their ambient levels, investigate and evaluate the sources of P-PAHs in the ambient air of Beirut.

**J-Sampling site:**

The sampling site is boarded by the Mediterranean coastal road 100 m west, by bliss street 200 m east and it is 4.2 Km far from Beirut harbor. The sampling site is surrounded from all sides by campus park and mostly pedestrian small roads (Figure8). All these criteria made our sampling site a wide site relevant for the study of the seasonal variation and for the evaluation of several sources into the ambient PAHs.





**Figure 8.** Location of the sampling site

## Chapter II

### P-PAH SAMPLING, ANALYTICAL METHOD OPTIMIZATION AND VALIDATION

In this chapter, a full description of the adopted sampling method as well as the optimized extraction and quantification procedure for the assessment of PPAHs is presented. Analysis of P-PAHs levels undergo three crucial stages: collection, sample preparation, and instrumental analysis. Samples of ambient air are collected using either a LVS or a HVS for comparison. Sample preparation include filter extraction, sample clean up, and sample concentration. The final stage is quantification of PPAHs, which is carried out using Gas chromatography-Mass spectrometry (GC-MS).

#### **A-PPAHs collection techniques: High and Low Volume Sampling Comparison**

EPA developed a compendium method TO-13 for the determination of PAH in ambient air using the high-volume sampling technique (Figure 9). In brief, the gas and particle phase fraction are trapped in series on a Polyurethane foam (PUF) (sorbent) and a 147mm quartz fiber filter, respectively, at a flow rate of 500 L/min. Both PUF and the filter are extracted using Soxhlet extractor and the analytes of interest are then isolated from the matrix using solid phase extraction (SPE), concentrated and finally analysed by GC/MS. The extraction method lasts for 16 hours and consumes up to 500 mL of hexane/diethylether as an extraction solvent for a single extraction at a time. However, some studies reported the use of low volume sampling (LVS) technique (Figure10). It is a collection method that uses lower flow rates (such as 10L/min) to collect PAHs on a 47mm quartz fiber filter. The LVS

technique uses Ultra sonication to extract PAHs, which last for 2 hours and requires much lower extraction solvent amount than the HVS extraction technique.

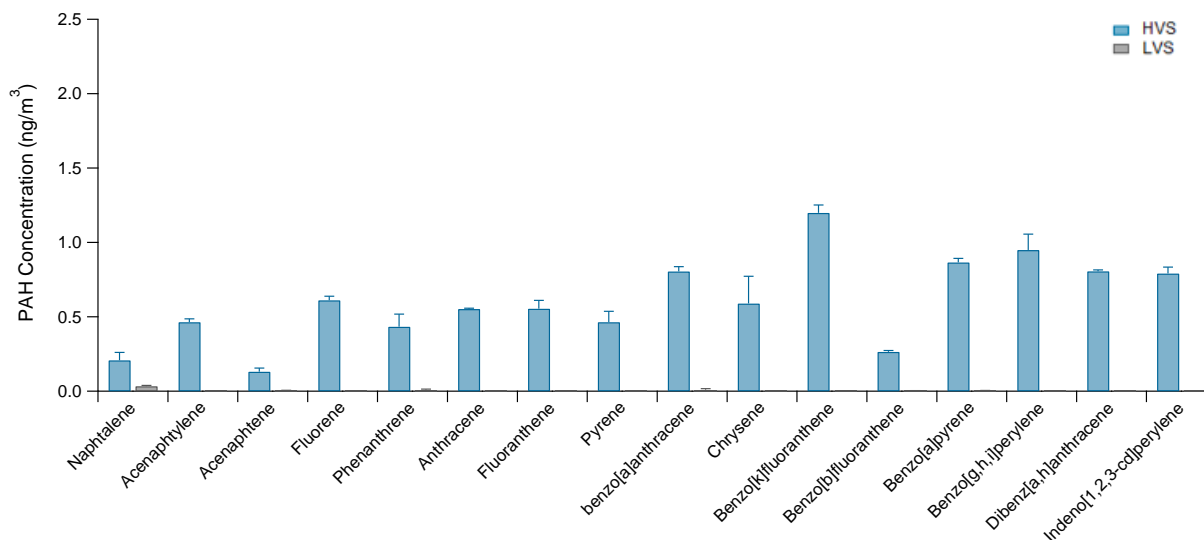
In order to test whether an LVS technique works with a background site like AUB, a low volume sampler (chemcomb Model 3500 Speciation Sampling Cartridge, Thermo electron Corp., Ohio, µUSA) operating at 10 L/min was run side by side for 24-hr with a high-volume sampler (DIGITEL enviro-sense DH77) operating at 500L/min. Both sampler's impactors are designed with a cut point to collect particles of an aerodynamic size of 10 µm in diameter. Prior to sampling, quartz-fiber filters were usually baked in a furnace oven at 500 °C for 5 hours to removed adsorbed organic compounds. Post sampling, the two filters were subjected to extraction and analysis. PAHs collected by the low-volume sampler at AUB were found to be below our detection limit. Other trials, which were repeated using composites of two and three low volume filters, were also unsuccessful in detecting PAHs. Figure 11 shows the comparison between a composite of three low volume filters and a high volume one. This issue was also encountered with Daher et al, where they were able to determine PAHs at the road site but not at AUB[86]. Hence, for this study, the high-volume sampling technique will be adopted but with an alternative optimized extraction method.



**Figure 9.** From left to right: the high-volume sampler, the PM<sub>10</sub> sampler impactor, and the HVS compartments constituting it



**Figure 10.** From left to right: the low volume sampler with a PM<sub>10</sub> impactor, and the setup



**Figure 11.** Comparison of PPAH concentrations between high and low volume sampling

## B-Sample preparation of HVS filters

### 1. Materials

PAH standards and deuterated internal standards containing 17 and 4 PAH compounds respectively, and dissolved in methylene chloride were obtained from Absolute standards. HPLC-grade toluene and hexane solvents were procured from sigma Aldrich. SPE cartridges (1000 mg/6 mL HyperSep SI) and quartz filters (Advantec, QR-100, 150 mm) were obtained from Thermo Scientific and Whatman International, respectively.

### 2. Filter extraction

In recent years, there has been a growing effort to reduce or to eliminate solvent use when extracting analytes of interest from various matrices and hence limit the impact of waste solvent on the environment. An extraction procedure was developed by optimizing four parameters: choice of solvent, extraction volume, extraction time and extraction

temperature. PAHs are known to be non-polar compounds and soluble in many solvents among which benzene, toluene, and dichloromethane have similar polarity to PAHs and are compatible with GC-MS. In the ideal case, benzene is the most suitable extraction solvent but due to its hazardous properties it was not used. Dichloromethane is more polar and volatile than toluene. Toluene is a nonpolar solvent that has a similar structure and compatible polarity to PAHs and is not a highly volatile organic solvent that cannot evaporate during extraction procedure and decreases the recovery of the analytes, for these reasons toluene was chosen as an extraction solvent for PAHs. To evaluate the extraction parameters, triplets of quartz filters were spiked with PAH standard and internal standard of 2 µg/ml. Each three replicate solutions were sonicated at 40°C for either 0.5, 1, and 1.5 hours and finally injected into GC-MS. The best recoveries of PAHs were for those sonicated for 1.5 hours which marked a recovery above 80%. A volume of 250 mL was found to be sufficient to extract PAHs without causing an excessive dilution and decreasing the waste solvent in comparison to Soxhlet which uses up to 500 ml of extraction solvent.

### ***3. Sample concentration***

Sample concentration is a critical requirement to increase PAH concentrations. Our aim is to concentrate the sample down to 1 mL. Rotary evaporation was employed because the sample of 250 ml would require a fast and an efficient concentration method of evaporation. Nitrogen evaporator was not considered because it is time consuming. Hence, a rotary evaporation was considered and the samples were concentrated at 45°C down to 1 ml.

#### ***4. Solid phase extraction***

In order to minimize the matrix interference, a clean-up procedure that purifies the sample from residual compounds other than PAHs is necessary. There are different types of clean-up techniques among which solid-phase extraction is the most applicable. The general procedure starts by conditioning the SPE cartridge, loading the sample and then eluting analytes of interest. The conditioning step was performed by washing the cartridge with 10 mL of hexane. This step is recommended by manufacturer 's instructions to activate the functional groups on silica surfaces. Subsequently the concentrated sample of 1 ml was loaded onto the SPE cartridge and finally elution was performed using 10 ml of hexane as a mobile phase.

#### ***5. Final concentration step***

After completing SPE procedure, a nitrogen evaporation step is essential to concentrate the sample for better precision in the quantification of the desired analytes. This step is crucial because PAHs are expected to be present in low ppm amounts in ambient air. A concentration volume down to 200  $\mu\text{L}$  was good to quantify PAHs with precision without over killing the final sample. Both the flow rate and the temperature of evaporation were investigated. The vapor pressures at 25  $^{\circ}\text{C}$  of the 16 PAHs ranges between  $2.1 \times 10^{-11}$  and  $8.9 \times 10^{-2}$  Pa and that for toluene is 3.8 Kpa. Consequently, sample concentration was carried out at atmospheric room temperature where PAHs are relatively less volatile than toluene. To determine the optimal flow rates, standard solutions of 2  $\mu\text{g/mL}$  were concentrated down to 200  $\mu\text{L}$  at 15 and 10 L/min. The recovery at 15 L/min was found to be between 60 to 90% for PAHs compared to greater than 80% for 10 L/min. Consequently, the slowest



flow rate of 10 L/min was selected. Having optimized all the sample preparation procedure, the sample is ready for GC-MS analysis.

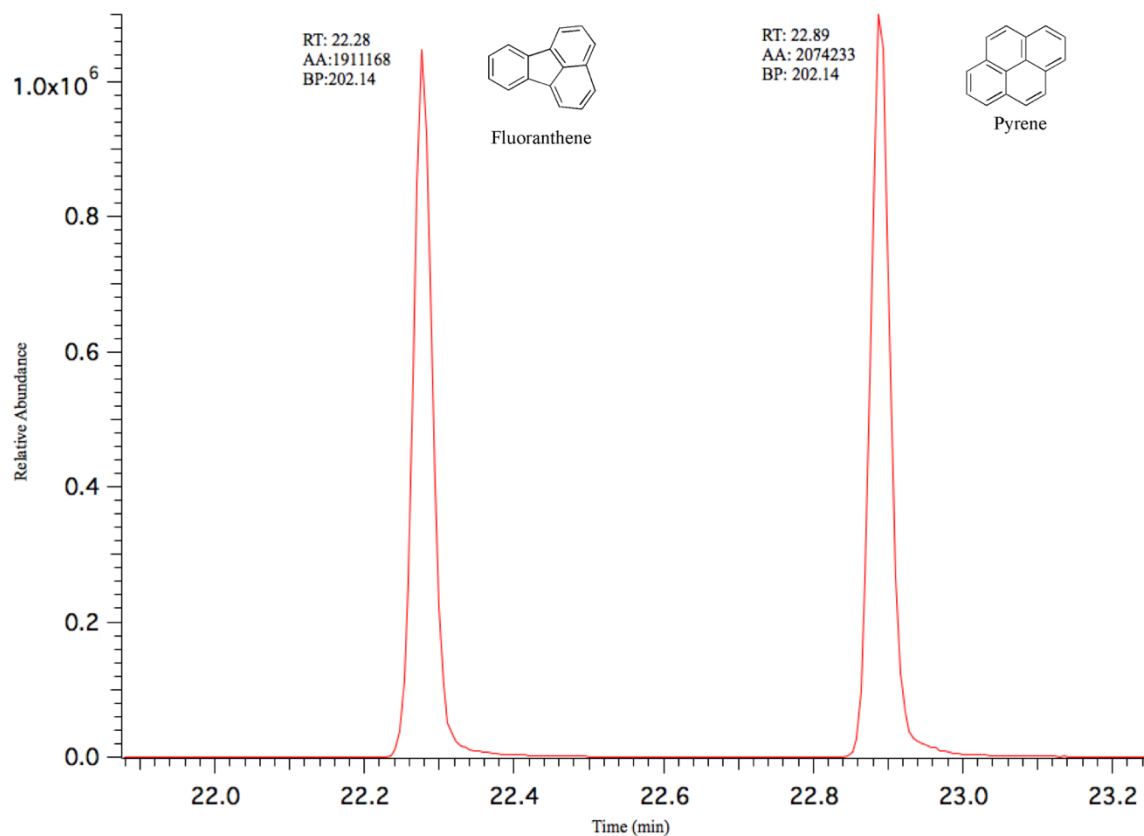
### **C-Instrumental analysis using GC-MS**

The analysis of PPAHs was accomplished by Thermo-Finnigan Trace GC-Ultra Polaris ITQ 900 MS coupled with AS 3000 II autosampler. Chromatographic separation was carried out on an Rtx-5MS column (60 m × 0.25 µm film thickness × 0.25 mm film ID). The carrier gas was helium of 99.999% purity with 1 mL/min flow rate. Injection mode was splitless and set at 280 °C. The GC oven was programmed from 80 °C (hold for 3 minute) to 170 °C (10 °C/min, hold for 1 min), to 180 °C (3 °C /min, hold for 0 min), to 270 °C (10 °C/min, hold time 0 min), then ramped to 300 °C (3 °C/min, hold for 10 min). The mass spectrometer was operated in full scan mode (50-350). The ion source temperature was 250 °C in electron impact mode (70 eV). The analytes were identified by their mass spectrum in which PAHs and IS have a relatively intense molecular ion (mass-to-charge ratio (m/z)). The linearity was evaluated by building an 8-point calibration curve in the range from 0.1 to 10 µg/mL, good linearity was observed with correlation coefficients ( $R^2$ ) > 0.995 for most of the 16 PAHs (see figures 12 and 13).

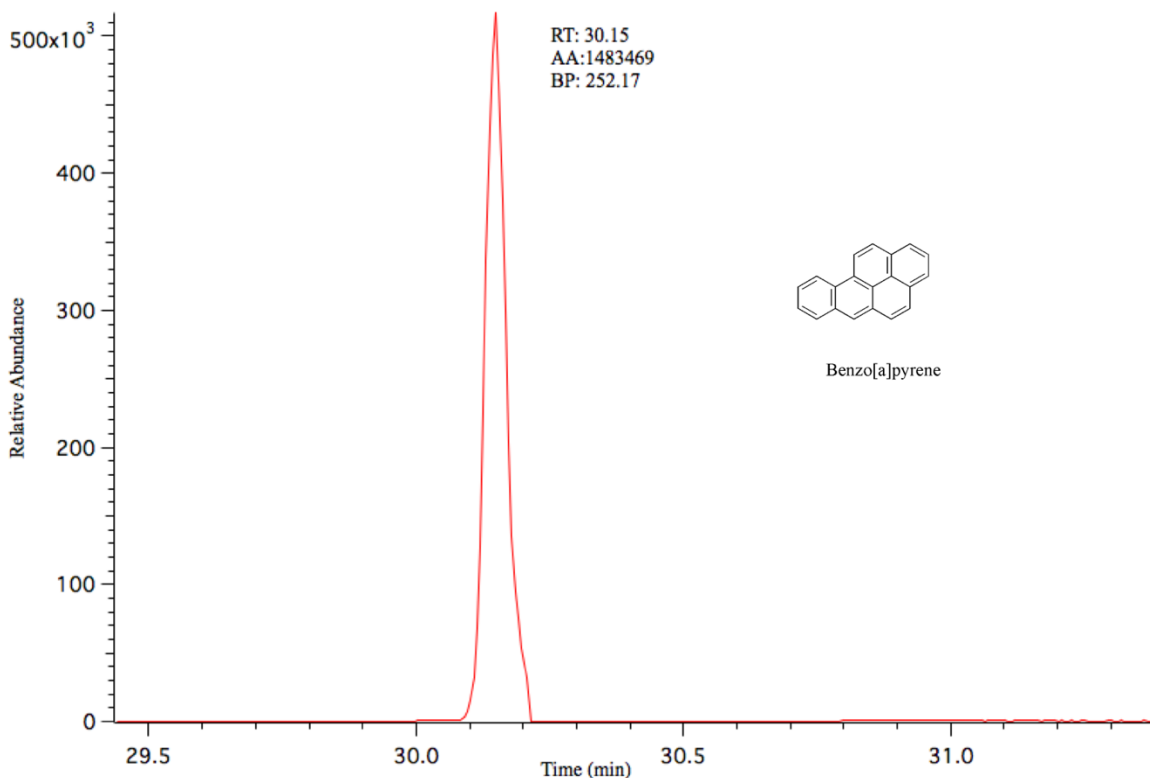
### **D- Quantification**

Using PAH reference standards containing 16 PAH compounds at concentrations ranging between 0.1 and 10 µg/ml and spiked by 2 µg/ml of deuterated PAH internal standard (IS) containing four PAH compounds which are acenaphthylene-d, phenanthrene-d, chrysene-d, perylene-d used to obtain the relative response factor for individual PAHs, an eight-points calibration curve was prepared. Quantification is based on a regression equation  $y = ax + b$  where  $y = \text{Area} [\text{standard}] / \text{Area}[\text{IS}]$ ,  $x$  is the concentration  $a$ ,  $b$  are the slope and intercept

respectively. Correlation factor ( $R^2$ ) ranged between 0.995 and 0.999 for the different PAHs calibration curves. The unknown concentration ( $x$ ) was computed as follows:  $x = y' - b / a$  where  $a$ ,  $b$  and  $y'$  are the slope, the intercept and  $Area[unknown] / Area[IS]$ , respectively.



**Figure 12:** Examples of the elution peaks for fluoranthene and pyrene of same molecular weight



**Figure 13.** Example of elution peak of benzo(a)pyrene the group one carcinogen among the PAH family.

### **E-Method validation**

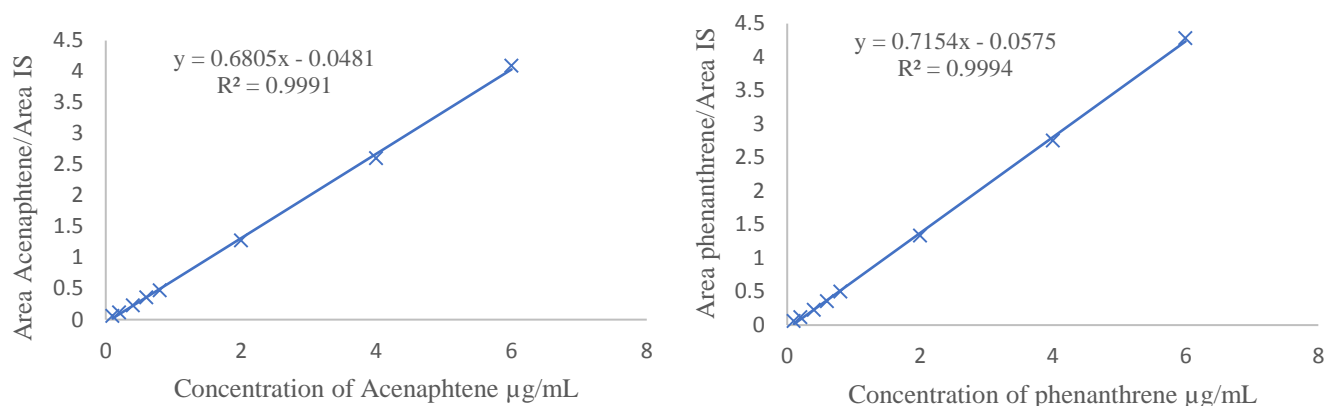
Validation is an integral part of quality control (QC) and quality assurance (QA) practices to judge the reliability, consistency and accuracy of the method. The key criteria for this evaluation are: linearity, limit of detection, limit of quantification, recovery and repeatability.

#### ***1. Linearity***

Linearity evaluation verifies that PAHs are found in a range where their response is linearly proportional to their concentration. It is commonly judged by examining the correlation coefficient of calibration curve. In this study, a direct calibration curve is used. PAH

standards ranging between 0.1 and 10 µg/mL are prepared and spiked with a constant amount of deuterated internal standards (2µg/mL)

The relationship between the ratio of the analytes signal to the IS signal and analyte standard concentrations was found to be linear for the whole examined range of the 16 PAHs with correlation coefficient >0.995 (Figure14).



**Figure 14.** Examples of direct calibration curves of Acenaphthene and phenanthrene

## 2. Method Limit of detection

Limit of detection (LOD) is the lowest concentration of analyte that can be detected but not necessary quantified. Based on the standard deviation response and slope, LOD is expressed as following:

$$\text{LOD} = \frac{3\sigma}{s}$$

Where  $\sigma$  is the standard deviation of the ratio of the analytes signal to the IS signal of seven replicates of analytes prepared at a low concentration and  $s$  is the slope of the direct calibration curve. LOD analysis was carried out using seven replicate extractions of 0.1

µg/mL spiked with 2 µg/mL of deuterated internal standard. The results have shown that detection limits of the 16 PAHs ranges between 0.01 and 0.05 µg/ml

### **3. Method Limit of Quantification**

Limit of quantification (LOQ) is the lowest concentration of analyte that can be measured with an acceptable level of accuracy and precision. Based on the standard deviation response and slope, LOQ is expressed as following:

$$LOQ = \frac{10\sigma}{S}$$

The quantification limits of the 16 PAHs were analyzed using seven replicate extractions of 0.1 µg/mL. The results have shown that LOQ ranges between 0.03 and 0.16 µg/ml

### **4. Repeatability**

Repeatability describes the closeness of agreement between a series of measurements obtained under the same operating conditions (one operator, same equipment and on the same day). It is expressed by the percent relative standard deviation (%RSD) of analytical results obtained from a minimum of five measurements at three different concentrations (low, medium and high). The acceptance criteria are based on type of analysis, complexity of matrix and the level of tested concentration. The results of six replicate standards for three concentrations (0.1, 4 and 8 µg/mL) revealed %RSD ranges between of 4 and 8%.

### **5. Recovery**

A valid extraction should reveal a high % recovery of the analytes. The recovery is the ratio of extracted concentration obtained from the sample treated according to the whole extraction procedure to that of a sample of same concentration directly analyzed on GC-MS. It is assessed using six extraction measurements over three concentration levels

covering the working range: 0.5, 6 and 10  $\mu\text{g/mL}$ . The recovery was found in the acceptable range between 83 and 110%. Consequently, all the QC and QA requirements are fulfilled and the method is ready for measuring PPAHs in ambient air.

#### ***6. Blank repeatability***

Field blanks, which accompanied samples to the sampling sites, were used to determine any background contamination using the same analysis method with samples. Concentrations of PAHs in the blanks were below the method detection limits in all the analysis period.

## Chapter III

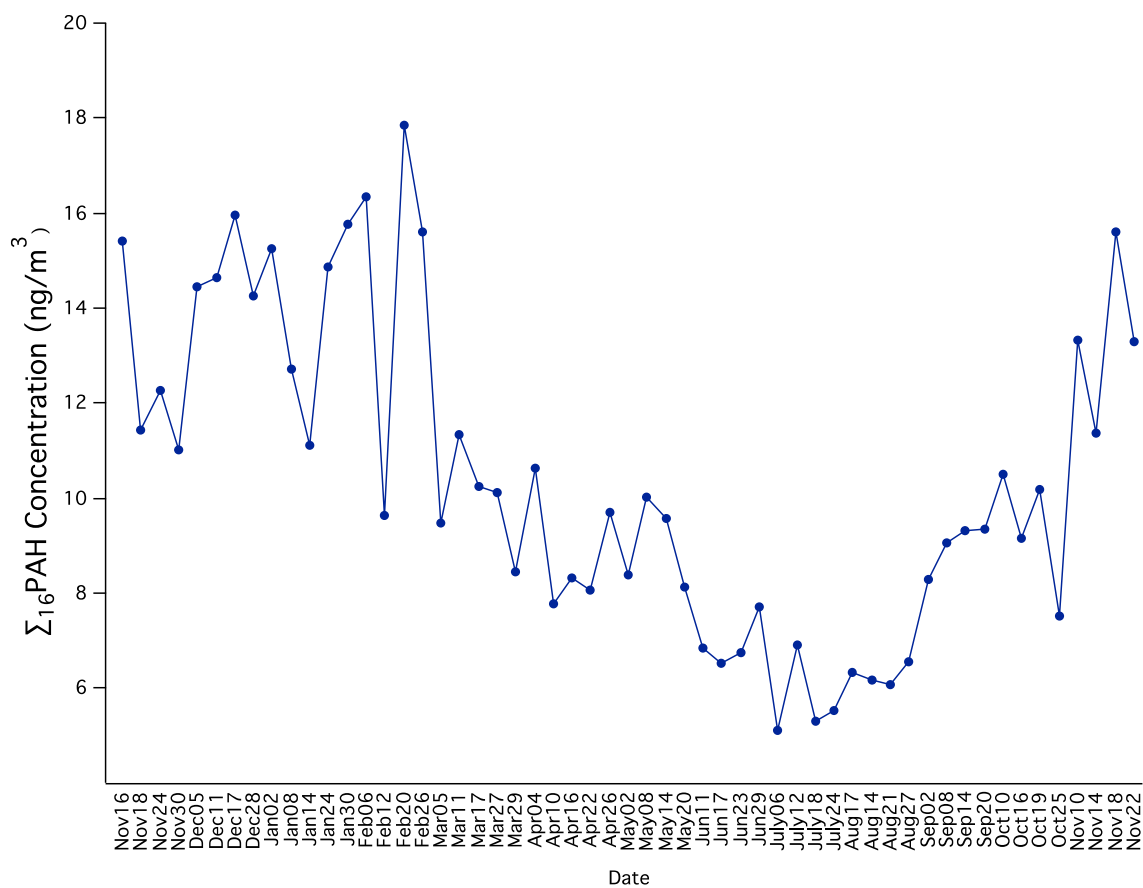
### RESULTS AND DISCUSSION

#### TEMPORAL AND SEASONAL VARIATION OF P-PAHs AT AUB

In brief, this work developed a method for the quantification of P-PAHs. After the method was validated, every six days P-PAHs were sampled using a high-volume sampler, extracted from filters and finally analyzed using GC/MS. The aims of this work after developing the method is to construct a baseline level of P-PAHs at an urban background site. Moreover, to monitor the seasonal as well as the intermonth variation of P-PAHs and to investigate the factors affecting their levels. Source apportionment was conducted to explore the main sources contributing to P-PAHs in the ambient air of an urban site. Toxicity evaluation is provided and incremental life time cancer risk was computed. Results are shown below.

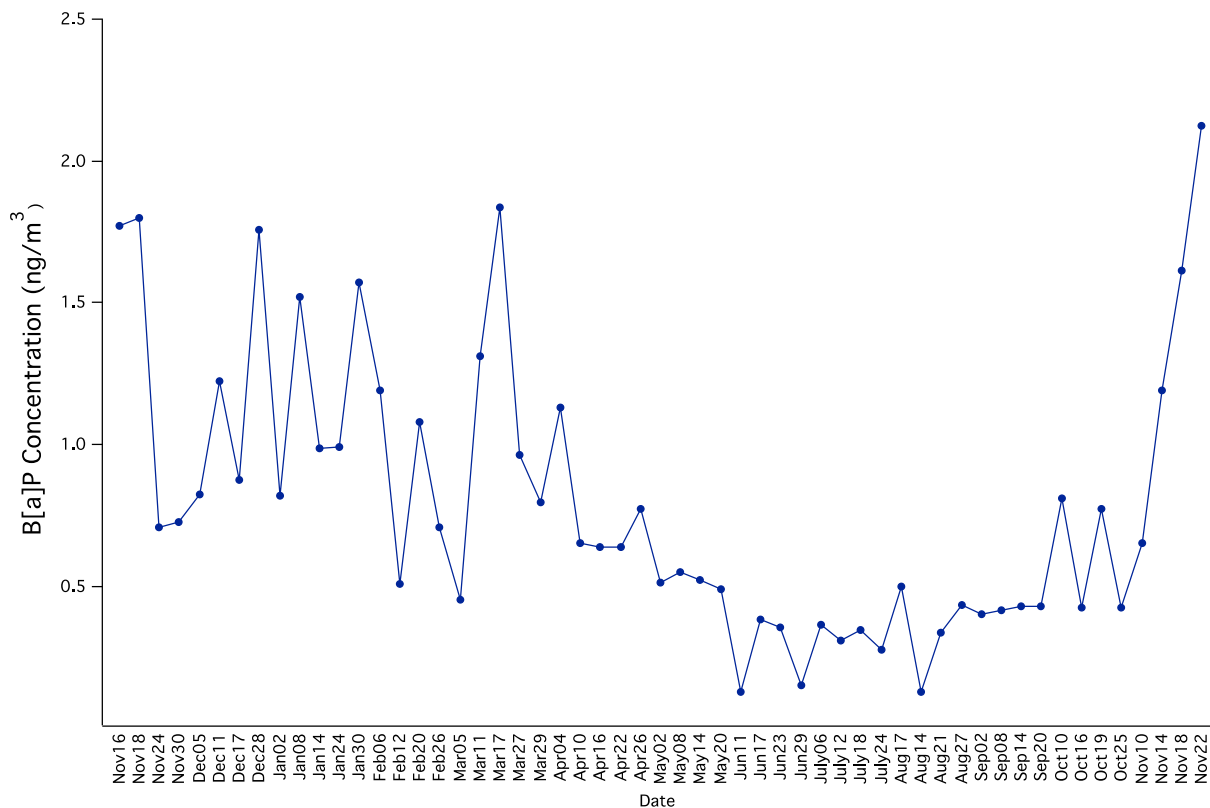
#### **A- variation of $\Sigma$ P-PAHs, P-B[a]p and individual P-PAHs**

##### **1-Temporal variation of $\Sigma$ P-PAHs, P-B[a]p:**



**Figure 15.** Temporal variation of the sum of P-PAH concentration on the sampled days





**Figure 16.** Temporal variation of B[a]P (Group 1 carcinogen) concentration on the sampled days

The data shows a high temporal variation throughout the year with higher concentration during winter and fall( Figure 15 and 16). The maximum sum of P-PAHs of 17.89 ng/m<sup>3</sup> was measured on February 20/2017 and the lowest of 5.1 ng/m<sup>3</sup> measured on July/06/2017. The high difference in concentration can be attributed to high variation in the meteorological conditions. For example, on February 20,2017 parameters such as temperature (14°C), average humidity (38%) and wind speed (1.66mps) favored a concentration of P-PAH higher than what was observed on November 24 under 20°C, 17% RH, and 2.77mps. Variations in meteorological conditions are suspected to have an effect

on the days that were sampled throughout the year. Of particular interest is a group 1 carcinogen, BaP, which followed the same trend of temporal variation during the sampled days. The concentration of B[a]P exceeded  $1\text{ ng/m}^3$  16 days out of 55 that were sampled. Most of the exceedances were during the winter and fall. The higher concentration detected for B[a]p was on November 18/2017 striking  $2.12\text{ ng/m}^3$ , whereas the lower concentration detected of  $0.13\text{ ng/m}^3$  was recorded on August 14, 2017.

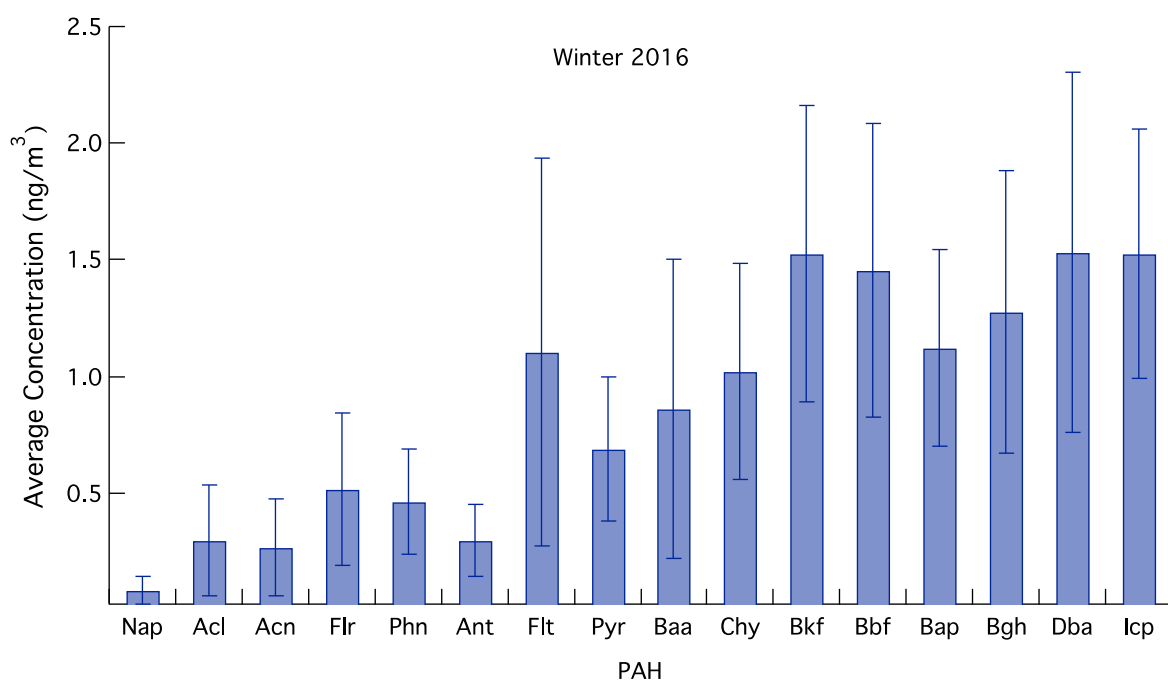
## **2-Seasonal variation of individual P-PAHs**

Figures 17-20 show the seasonal variation of the individual P-PAHs during the four seasons. A strong seasonal variation of P-PAHs was observed. During the winter and the fall seasons, the standard deviation of the individual P-PAHs was high. This high deviation is attributed to the unstable wind speed during cold period which can either induce the accumulation or dispersion of P-PAHs. Figure 21 shows the effect of wind speed during the cold period on P-PAHs sum. There is a negative correlation between the P-PAHs and wind speed which in turn has an effect on increasing the standard deviation. However, this deviation decreases during spring and summer where the weather and specifically the wind speed is more stagnant and stable.

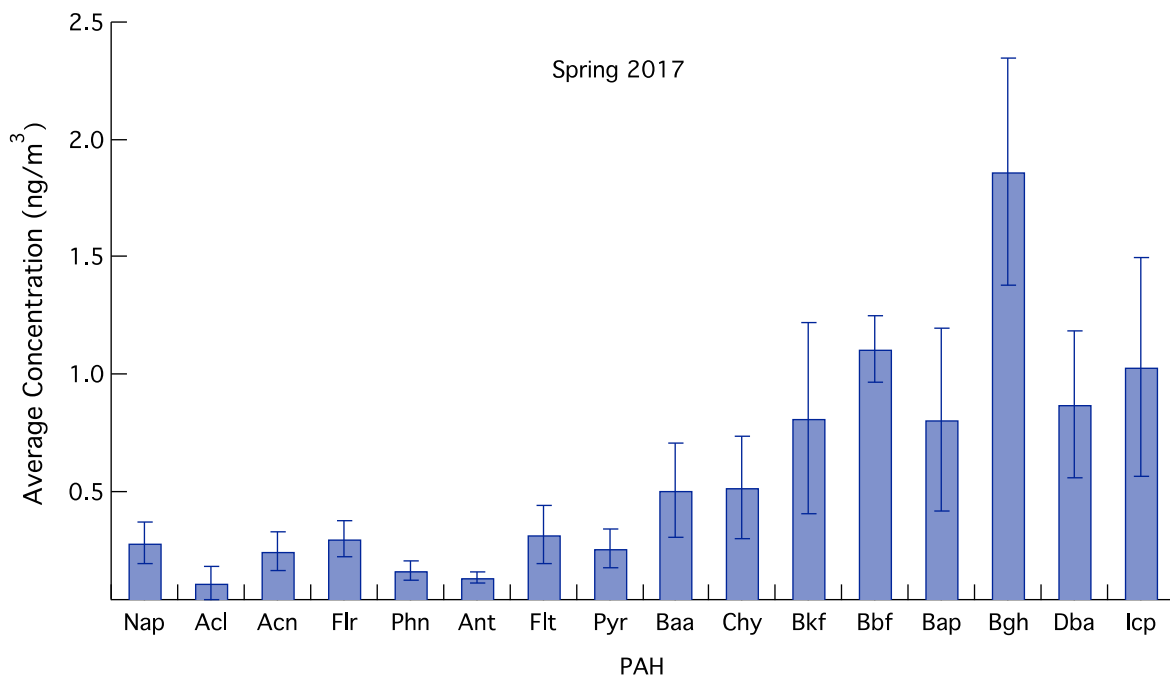
It is clear that as the molecular weight increase, the PAHs are more prone to partition to the particle phase. In fact, this is the case of Benzo[a]anthracene, Chrysene, Benzo[k]flouranthene, Benzo[b]flouranthene, Benzo[a]pyrene, Benzo[ghi]perylene, Dibenzo[a,h]anthracene, Indeno[1,2,3-cd]pyrene, characterized with a low vapour pressure ( $V_p < 10^{-6}\text{ Pa}$ ) and almost found in the particle phase. However, Naphthalene, Acenaphtylene, Acenaphtene, Flourene, Phenanthrene, Anthracene, Flouranthene and

Pyrene have a lower concentration due to their partitioning between the particle and the vapor phase, which is more prominent during the warm seasons that increases their vaporization rate ( $V_p > 10^{-6}$  Pa).

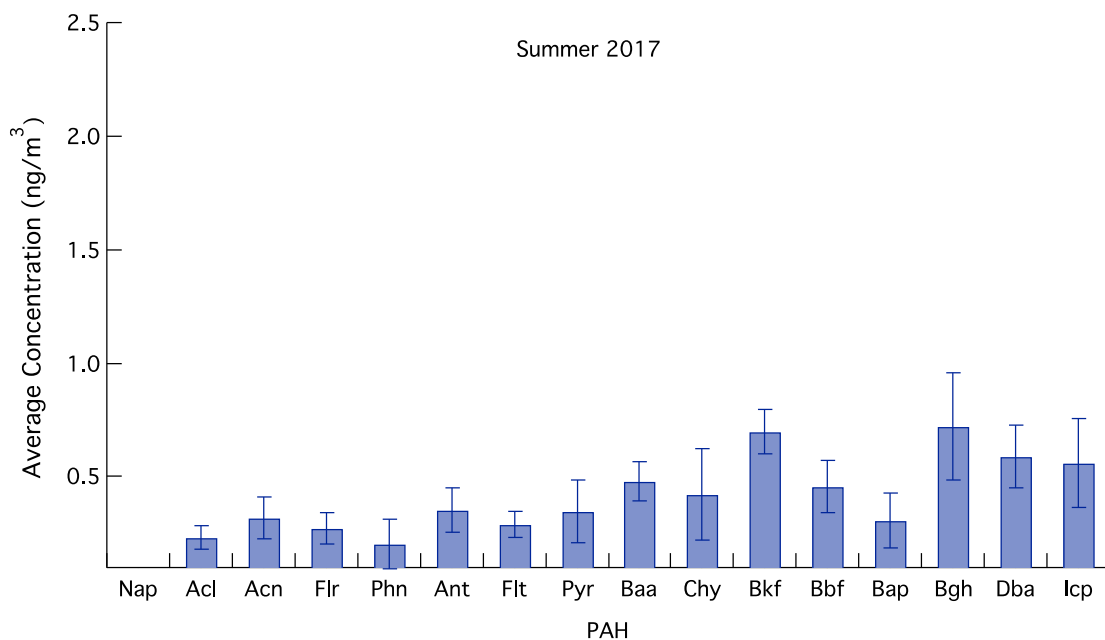
The results of this study are comparable to those of Teixeira et al.[91], which show higher HMW PAHs for the winter time. Thus, the magnitude of HMW PAHs was increased with increasing aerosols found in the winter time due to meteorological variables as well as the increased source emissions in the winter time.



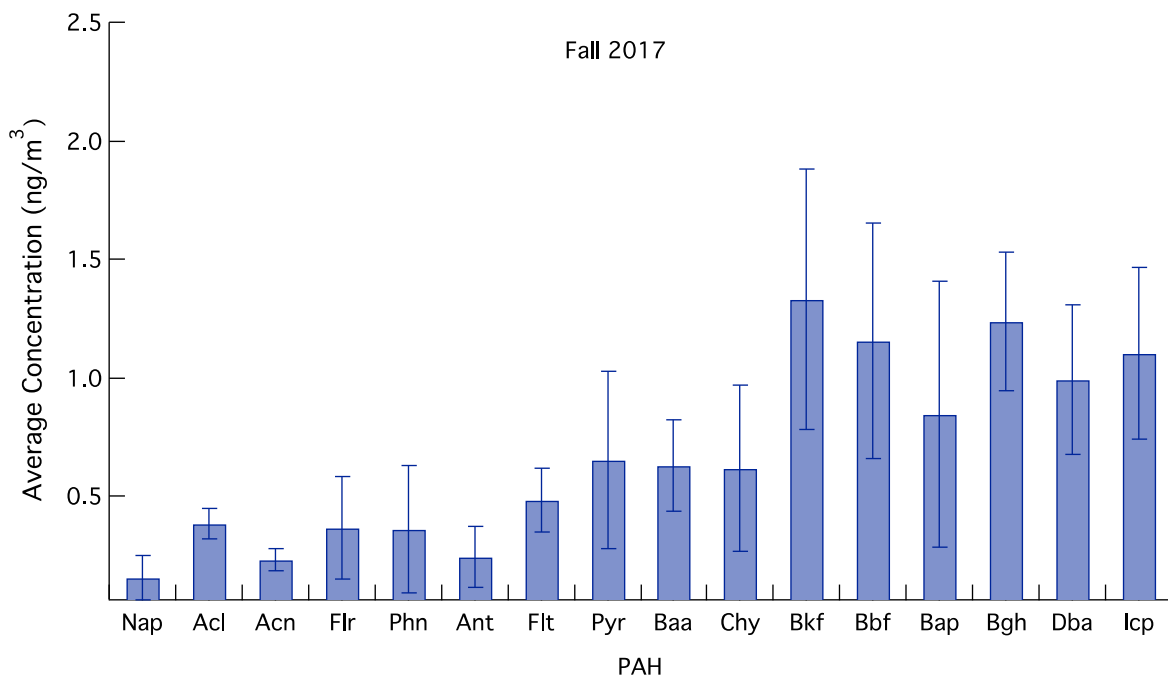
**Figure 17.** Seasonal variation for the average concentration of the 16 PAHs during the winter season extending between November 16, 2016 to February 26, 2017 (n=17)



**Figure 18.** Seasonal variation for the average concentration of the 16 PAHs during the spring season extending between March 5, 2017 to may 20, 2017 n=14

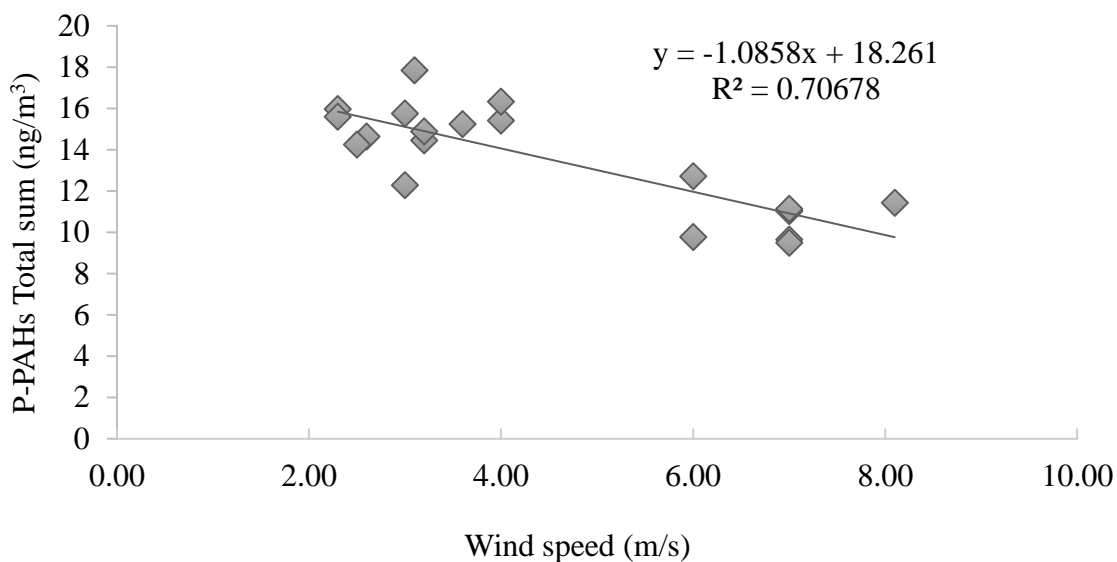


**Figure 19.** Seasonal variation for the average concentration of the 16 PAHs during the summer season extending between June 11, 2017 to august 27, 2017 (n=12)



**Figure 20.** Seasonal variation for the average concentration of the 16 PAHs during the Fall season extending between September 9, 2017 to November 22 2017 (n=12)

**Abbreviations:** Nap(Naphthalene), Acl(Acenaphtylene), Acn(Acenaphtene), Flr(Flourene), Phn(Phenanthrene), Ant(Anthracene), Flt(Flouranthene), Pyr(Pyrene), Baa(Benzo[a]anthracene), Chy(Chrysene), Bkf(Benzo[k]flouranthene), Bbf(Benzo[b]flouranthene), Bap(Benzo[a]pyrene), Bgh(Benzo[ghi]perylene), Dbal(Dibenzo[a,h]anthracene), Icp(Indeno[1,2,3-cd]pyrene)

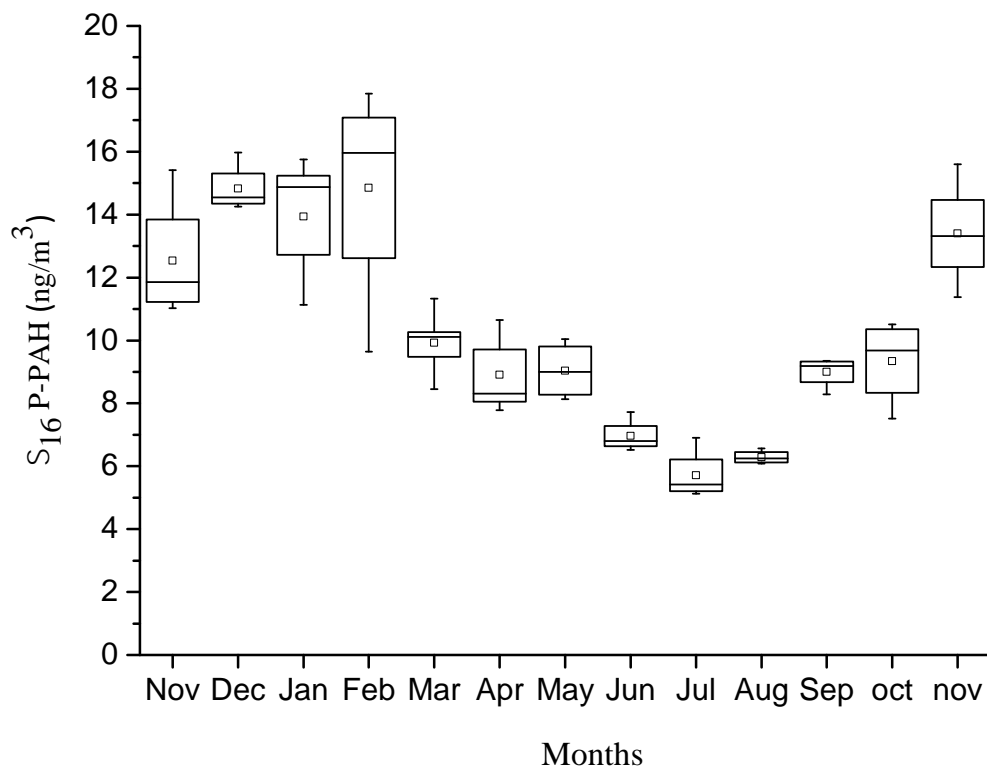


**Figure 21.** Correlation between sum of P-PAHs concentration and wind speed during the cold seasons

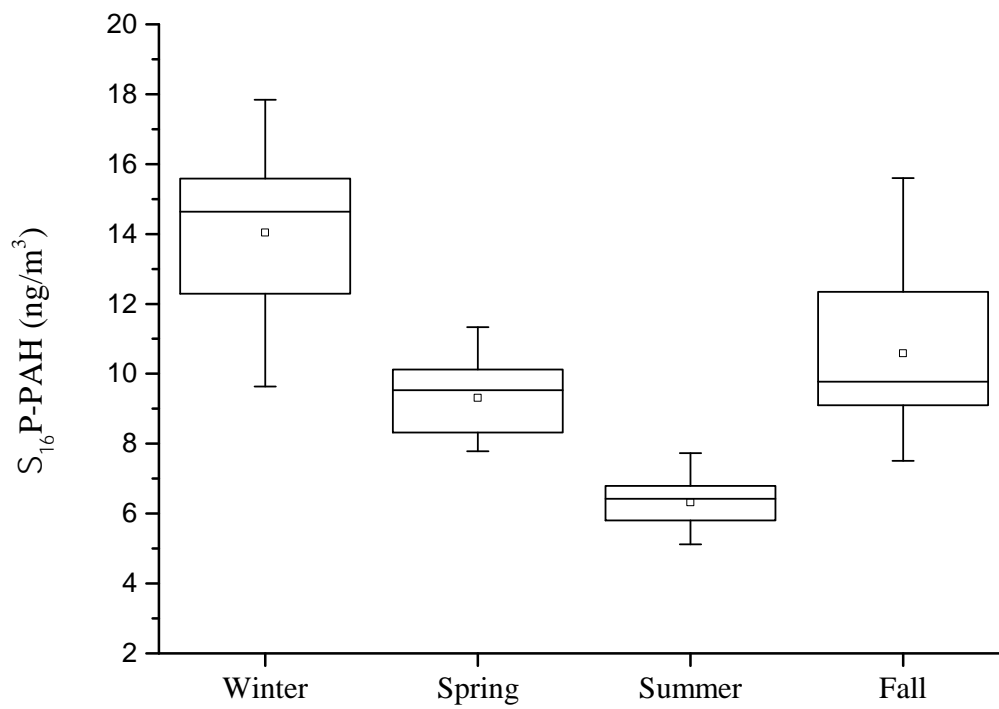
### 3-Intermonth and Seasonal variation of $\sum_{16}$ P-PAH and P-B[a]P

Whisker and box plots are shown in figures 22-25 for the P-PAH sum and P-B[a]p during the 12 months campaign and the four seasons. The whisker plots indicate how the points vary from each other during the month or the season. The small squares and the line inside

the box represent the mean and the median respectively.

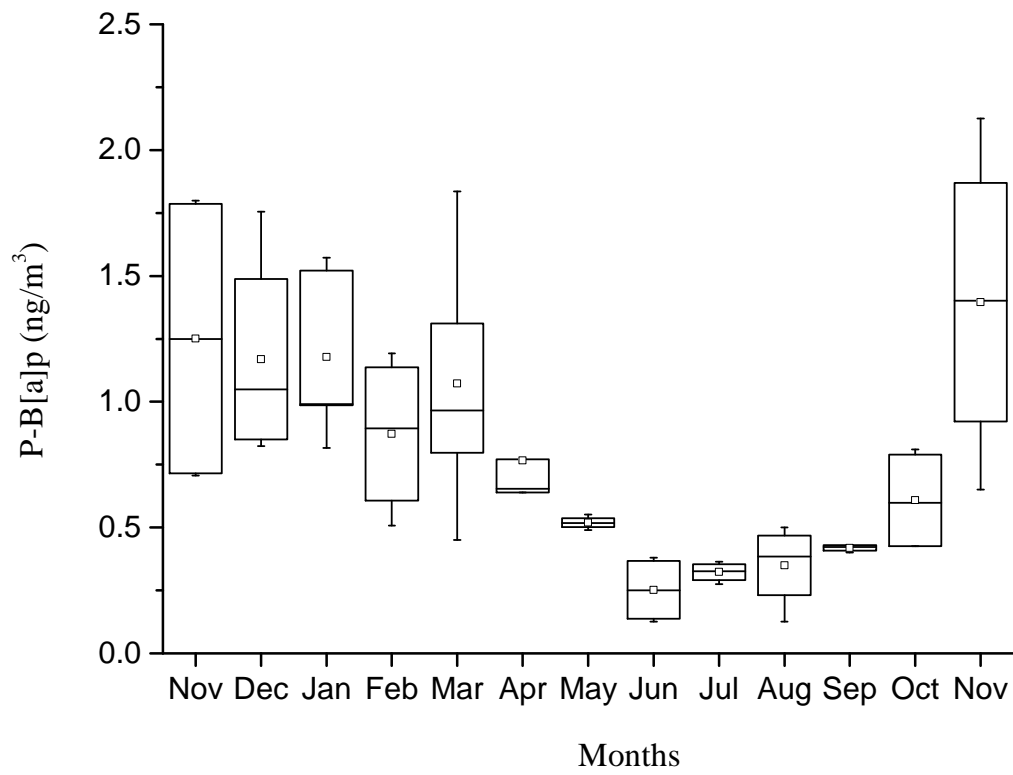


**Figure 22.** Whisker and box plot of  $\Sigma$ P-PAH during the twelve months campaign

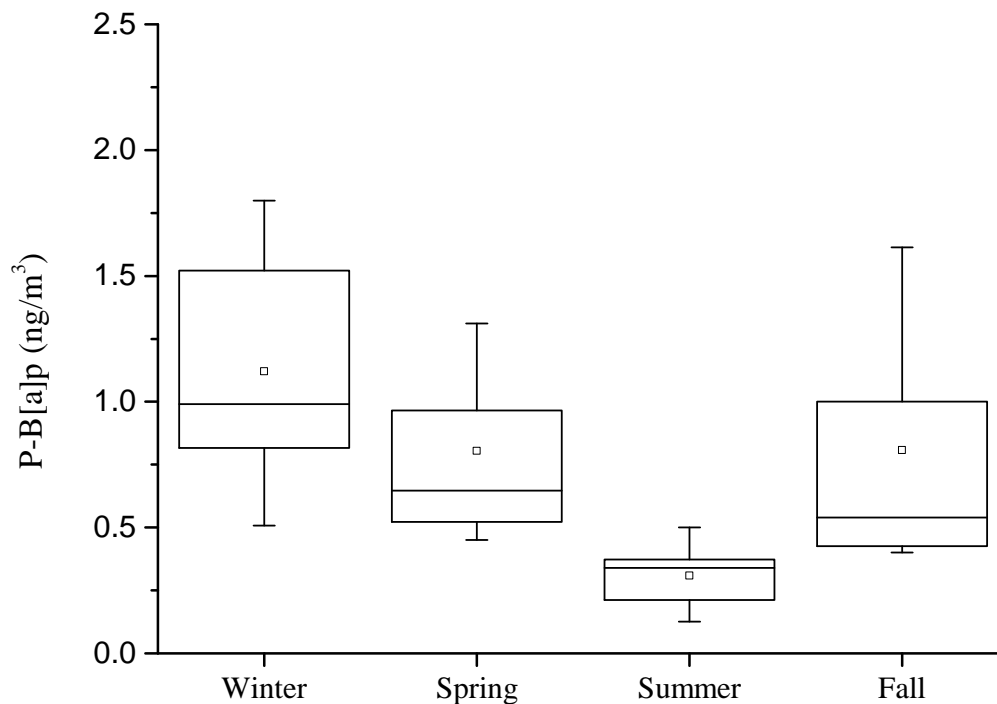


**Figure 23.** whisker and box plot of  $\Sigma$ PPAH during the four seasons





**Figure 24.** whisker and box plot of P-B[a]p during the twelve months campaign



**Figure 25.** whisker and box plot of P-B[a]p during the twelve the four seasons

The P-PAHs mean increased from  $12.9 \pm 2.0$  in November 2016 to  $14.9 \pm 3.0$  in February 2017. Then it started decreasing until reaching the lowest value of  $6.3 \pm 0.2$  in July 2016. Finally, P-PAHs started increasing gradually to reach  $13.4 \pm 1.7$  in November 2017. The mean of P-PAHs during the seasons was  $14.0 \pm 2.3$ ,  $9.3 \pm 1.1$ ,  $6.3 \pm 0.7$ ,  $10.8 \pm 2.4$  for winter, spring, summer, and fall respectively. The annual mean for the P-PAH sum was  $10.4 \pm 1.3$ .

P-B[a]p concentration fluctuated between  $1.3 \pm 0.6$  and  $1.1 \pm 0.52$  between November 2016 and March 2017, then the concentration decreased to  $0.3 \pm 0.2$  in August 2017. Finally, it increased again to reach  $1.4 \pm 0.6$  in November 2017. The mean B[a]p of the seasons was

1.1±0.4, 0.8±0.4, 0.3±0.1, 0.8±0.6 for winter, spring, summer and fall respectively. The annual mean for B[a]p was 0.8±0.3. Although the annual guideline value of B[a]P is 1.0 ng/m<sup>3</sup> according to Directive 2004/107/EC was not exceeded during this campaign, the UK Air Quality guideline value of 0.25 ng/m<sup>3</sup> was exceeded.

## **B-Discussion of Results:**

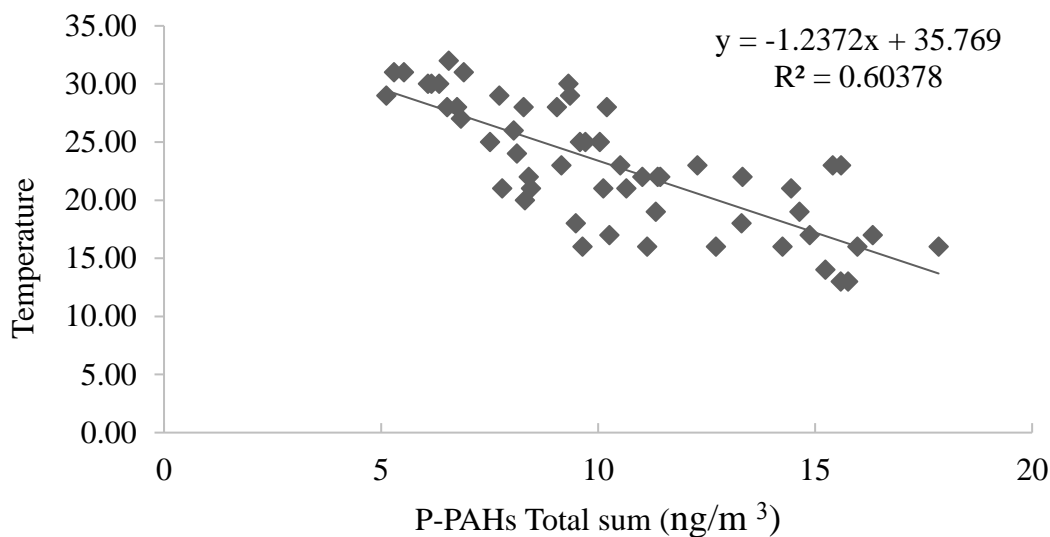
The variation of P-PAHs in air depends on the time of the year where both the seasonal emission sources and meteorology are different. The increase of P-PAHs in the winter and fall have been reported in many studies[92-97]. In the cold seasons, the increase in PAH concentrations can be related to the emission from domestic heating that are absent during the warm seasons[32, 98]. Furthermore, the low ambient temperature during this period has an effect on reducing the efficiency of fossil fuel combustion in the vehicular engines.

According to Devos et al, PAH emissions from gasoline vehicles are 10 times higher than that of diesel vehicles because of the cold starts of the engines during winter[29, 99-101].

The meteorology too has a great impact. One main aspect is the inversion layer which frequently occurs during the long cold winter nights where there is an excessive nocturnal cooling of the ground surface due to rapid rate of loss of heat from the ground that exceeds the amount of radiation received from the sun. This phenomenon restricts the upward vertical mixing and dispersion of PAHs and traps them below the inversion layer near a ground level [102-107]. The cold period is characterized with low ambient temperature, reduced solar radiation and high relative humidity which all limit the physical and chemical decomposition of P-PAHs, which in turn increases their persistence in the atmosphere[36,

105, 108-110]. Also, enhanced PAH condensation/adsorption to suspended particles often occurs at lower temperatures [29, 32]. Moreover, the size distribution of particles varied by season could also alter PAH concentrations. Duan et al.[111] have indicated that the accumulation mode of PAHs is slightly shifted to larger particles in cooler seasons as comparing with that in a warm season. The magnitude of aged particles with PAHs is decreased during warm seasons due to the presence of good dispersion conditions. It is likely that larger particles tend to be correlated with aged particles by Kelvin effect[112].

On the other hand, the high temperatures in the summer as well as the high solar radiation and low humidity favors the thermo-, photo- and chemical- oxidation for decomposing atmospheric P-PAHs. Furthermore, PAHs are known as semi volatile organic compounds and their distribution in the atmosphere between the gas and particle phases is highly influenced by the ambient temperature. Higher temperatures during the warm seasons yield a relatively larger portion of PAHs partitioned to the gas phase especially those of the low molecular weight. Figure 26 shows the correlation between temperature and P-PAHs sum. It is evident that as the temperature increase P-PAHs decrease.



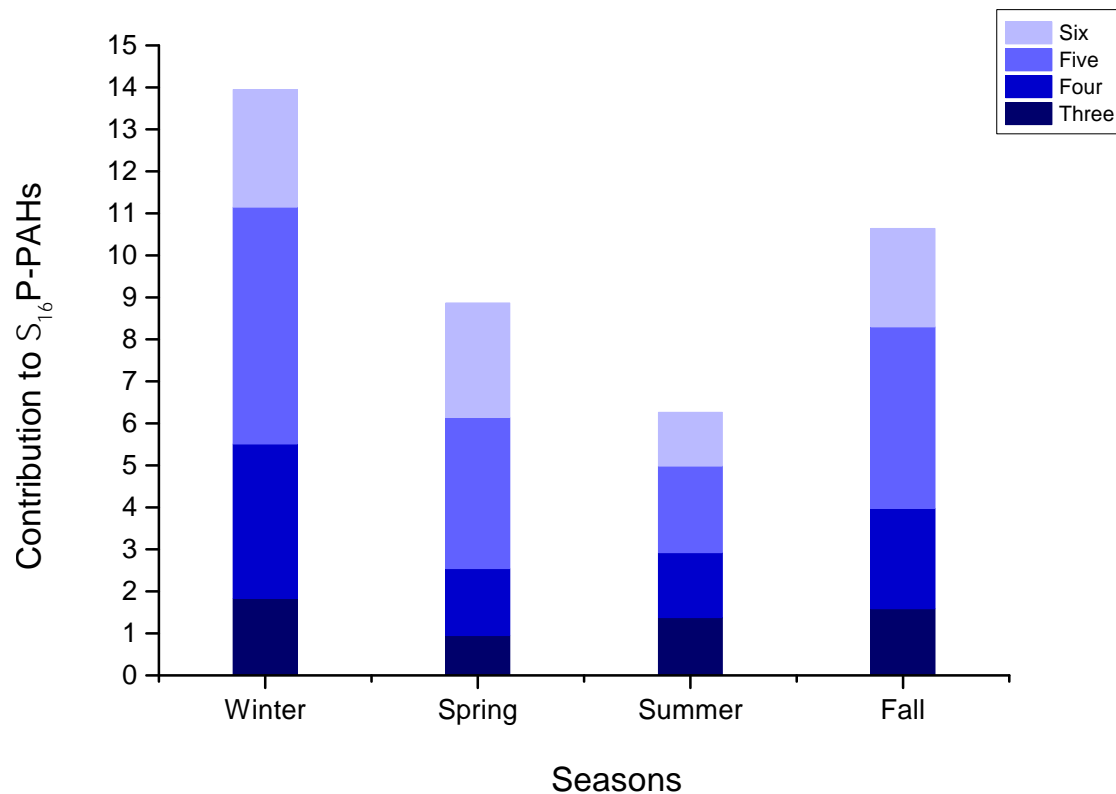
**Figure 26.** Correlation between sum of P-PAHs and temperature during the cold and warm seasons

### C- Classification of P-PAHs

Classification of P-PAHs can be done based on the number of their aromatic rings, and this was used to construct the distribution pattern of the P-PAHs (Figure 27) in each season to better understand their source emissions. The main components of P-PAHs in every season at all sites were those of the four to five aromatic ring that constitute Baa, Chy, Bkf, Bbf, Bap, Bgh, Dbf, Icp. These eight dominant components covered 55%–66% of the total 16 PAHs and exhibited a variation throughout the seasons of the year in contrast to the three or six member rings which didn't show a significant variation during the seasons. This indicates the effect of the increase in the emission sources for the four and five-member ring PAHs during the cold period as described before. These results suggest that the

emission sources and/or meteorological conditions in winter are quite different from those of the other seasons.

The combustion-derived PAHs (COMPAHs) have been classified by some authors [113-122], and include Fluo, Py, Chry, BbF, BkF, BaA, BeP, BaP, Indeo and BghiP. This criterion was applied to our data. The sum of the main combustion-derived PAHs concentrations (COMPAHs) covered 67%–77% of the total PAHs in PM<sub>10</sub>. In all seasons, the combustion-derived PAHs represented more than half of the total PAHs. Several studies determined that the ratio of  $\sum\text{COMPAH}/\sum\text{PAHs}$  is an indicator of the PAH origin whether they are of petrogenic or of pyrolytic origin.  $\sum\text{COMPAH}/\sum\text{PAHs} \leq 0.3$  indicates petrogenic origin while  $\sum\text{COMPAH}/\sum\text{PAHs} \geq 0.7$  indicates a pyrolytic origin [106, 123-127]. In this study, at AUB the value of  $\sum\text{COMPAH}/\sum\text{PAHs}$  at AUB was in the range of 0.67-0.77 throughout the seasons which highly indicates the effect of combustion sources (pyrogenic) such as diesel and gasoline combustion on the PAHs at AUB. Another ratio is considered to understand the fate of LMW PAHs which is the ratio of LPAH/HPAH. Several studies pointed out that the concentrations of LMW PAHs in ambient air might not reflect their origin because they are more susceptible to atmospheric degradation. A ratio  $0.2 < \text{LPAH}/\text{HPAH} < 0.4$  indicates that degradation of LPAH is likely to have occurred [128-130]. At AUB, the ratio was between 0.2 and 0.3 which indicates that high degradation of LMW PAHs have occurred.



**Figure 27.** Contribution of the three, four, five and six member rings to the total P-PAH sum.

#### **D- Bap Toxicity Equivalent concentration**

B[a]p is the most PAH known for its carcinogenic and mutagenic character. Nevertheless, tracking this PAH alone would underestimate the carcinogenic effects of the other PAHs. Accordingly, B[a]p toxicity equivalent concentration has emerged to focus on the total toxicity associated with the exposure to a mixture of PAHs. The individual carcinogenic potencies of PAH in relation to B[a]P can be expressed through the B[a]P equivalent

concentrations ( $BaP_{eq}$ ), by multiplying the concentration of each PAH by the respective TEF value, using the following equation:

$$BaP_{eq} = \sum_{i=1}^n C_i \times TEF_i$$

Where  $C_i$  is the concentration of PAH congener  $i$  and TEF is the toxicity equivalency factor of the PAH congener  $i$  relative to B[a]P.

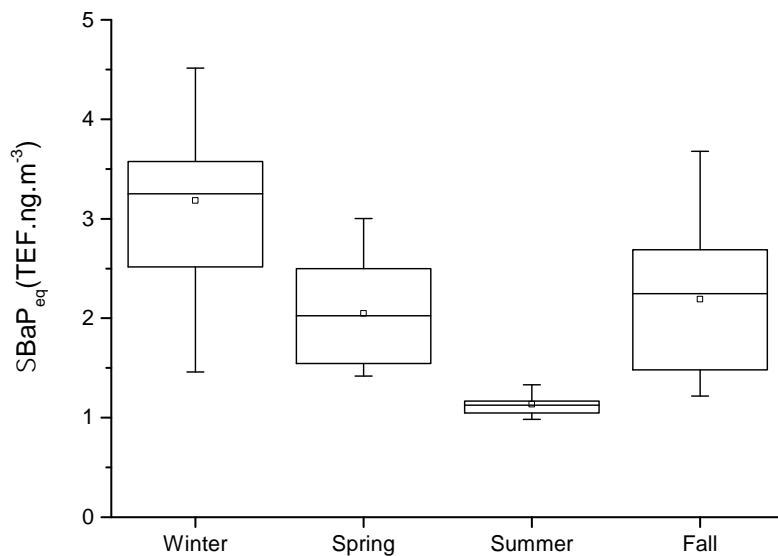
In this work, the TEF concept was applied and values of TEF reported by IARC were used to calculate the TEF-adjusted concentrations (based on BaP) of carcinogenic PAHs. The total toxic equivalent represents the result of the sum of the  $BaP_{eq}$  values ( $\sum BaP_{eq}$ ). The mean value obtained for the carcinogenic potential estimated through the sum of  $BaP_{eq}$  in the four seasons was  $3.2 \pm 0.9$ ,  $2.1 \pm$ ,  $1.2 \pm$  and  $2.3 \pm$  for winter, spring, summer and fall respectively. The annual mean for  $BaP_{eq}$  was  $2.1 \text{ ng TEF m}^{-3}$ . The results show that higher potential risk for human health upon exposure to the PAHs mixture is expected during the winter season( Figure 28).

The fraction between each  $BaP_{eq}$  and the total toxic equivalents ( $\sum BaP_{eq}$ ) provides the percentage contribution of each PAH to  $\sum BaP_{eq}$ . *Db*a, which was the specie found in high concentrations and due to its corresponding TEF (TEF of 1), was the highest contributor to  $\sum BaP_{eq}$ , with an average of 47.0 % for the period of 2017, followed by BaP with a 35.0 % contribution.

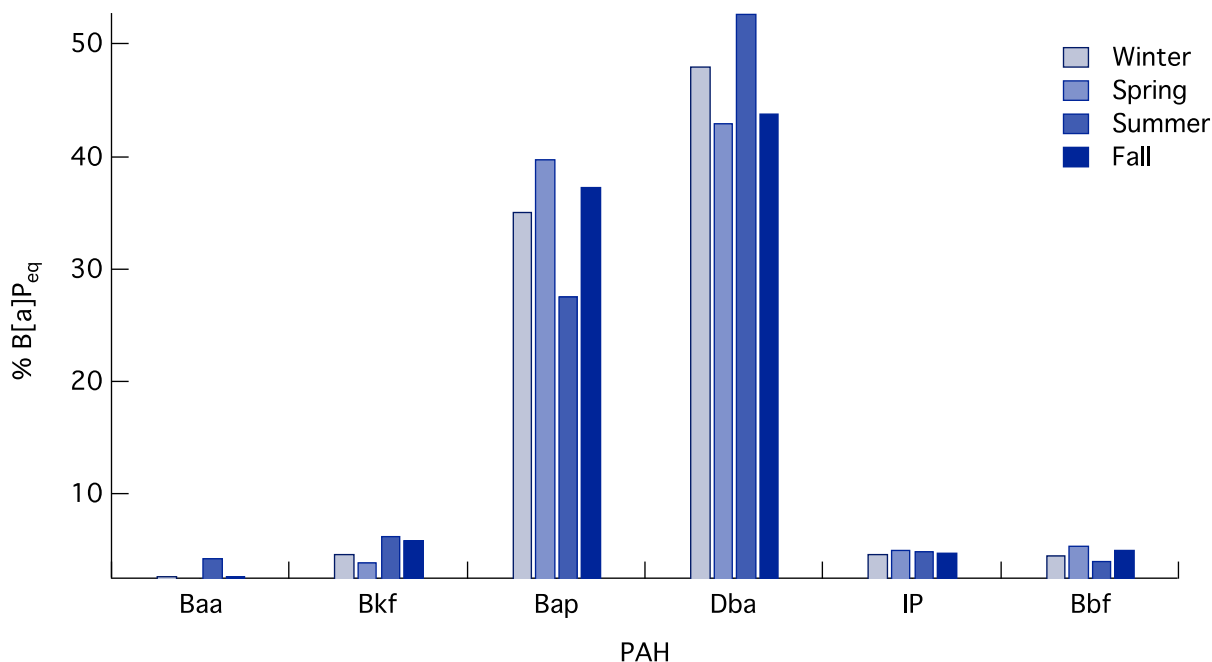
Of all the 16 PAHs considered priority pollutants by the US-EPA, only BaA, Chry, BaP, BbkjF, Ind and *Db*a were considered potential carcinogens [131] and, in this study, these compounds have been the species with the higher contribution in relation to  $\sum BaP_{eq}$ .



Figure 29 shows the equivalent toxicity concentration contribution for these six species in the four seasons. The contribution of carcinogenic potential of B[a]P ranged from ~ 27.6 (summer) to ~ 39.8% (spring) and Dba ranged from ~ 43.1% (spring) to ~ 52.8% (summer). The remaining species showed contributions below 10% to  $\sum \text{BaP}_{\text{eq}}$ . The high value obtained was related to a higher contribution from the Dba with an average toxicity concentration of  $0.996 \text{ ng TEF m}^{-3}$  in 2017. The results suggest that focusing only on B[a]P, which is mostly used as a marker of the genotoxic and carcinogenic PAH, would probably underestimate the carcinogenic potential of the other studied PAHs. It should be noted that Dba is listed in the group 2A carcinogen family with a TEF equals to 1 in that family.



**Figure 28.** Seasonal variation of B[a]p equivalent toxicity concentration



**Figure 29.** Percent contribution of the most carcinogenic PAHs to the B[a]P toxicity equivalence

### E-Incremental lifetime cancer risk assessment

A growing number of studies have highlighted the health risks of atmospheric PAHs, especially those on fine particle [132-135]. The metric method used for estimating cancer risk is the inhalation incremental lifetime cancer risk (ILCR), which is associated with specific inhalation and exposure parameters, and calculated from the following equation:

$$ILCR = \frac{E \times SF \times EF \times ED \times CF}{BW \times AT}$$

in which E is the daily exposure level by inhalation ( $\text{ng}\cdot\text{day}^{-1}$ ) for a specific age which is calculated using the toxic equivalent concentration sum and the inhalation rate (IR) for a specific age group ( $21.4 \text{ m}^3/\text{day}$  for adults) and the time of exposure which was taken as 23 hours using the following equation:  $E = \sum BaP_{eq} \times IR \times \text{Time of exposure}$

The ILCR can then be calculated. *SF* is the inhalation carcinogenic slope factor for BaP, which has a geometric mean of 3.14 kg.day/mg, and a geometric standard deviation of 1.8 [136], *ED* means the exposure duration (50 years for adults), *EF* is the exposure frequency (252 day. year<sup>-1</sup>), *BW* is the body weight (70 kg for adults) and *AT* is the lifespan of carcinogens (25,550 days); and *CF* (mg/ng) is a unit conversion factor (10<sup>-6</sup>)[137].

In our study, ILCRs for adult's population at the urban site, were estimated. This age was selected because it has the highest inhalation rate and exposure duration. A scenario of an adult individual living all time at the same level of the rooftop at AUB and exposed to this ambient level was simulated. It is worth noting here that the values obtained represent a lower limit exposure risk since only the inhalation route was assessed and other exposure routes (ingestion and dermal contact) were not considered.

The computed ILCRs from exposure to airborne PAHs were estimated assuming that adult lives in the urban site of the sampled area for 24 hours and for 365 days/year. Computed ILCR averaged at  $1.4 \times 10^{-6}$  in winter period which was the only significant value that exceeded the commonly acceptable threshold determined by IARC of  $10^{-6}$ . To sum up, these results show that out of a  $10^6$  population one adult individual would have the chance to develop cancer upon exposure to the PAH baseline mixture in the urban site. However, in daily life and at the exposure levels and not at rooftops ambient levels individuals are more exposed to PAHs and ultimately this value of cancer risk estimation would be higher.

## CHAPTER IV

### SOURCE APPORTIONMENT OF P-PAHs USING POSITIVE MATRIX FACTORIZATION

This chapter presents the major receptor models available for source apportionment. Among these models, we highlight the features of Positive matrix factorization (PMF), the model that is widely used to partition sources that contribute into air quality assessment. For this model to be valid, certain parameters should be first optimized. Therefore, these parameters are further detailed. Upon optimization, an interpretation of this model's output data can finally be generated.

#### **A-Introduction to receptor modeling:**

source apportionment using receptor modeling has been developed in order to elicit information about the sources that might be affecting a certain receptor site for a particular contaminant. The most widely used models are: chemical mass balance (CMB), principal component analysis (PCA), and positive matrix factorization (PMF)[138]. In brief, CMB is a regression based mass balance methodology proved by various studies to be efficient in modeling [139]. It assesses the percent contribution of a single source to the contaminant in a receptor site. The application of CMB is appropriate only when the source and its profile is known, so in this case the contribution of the source for a given data set is calculated. When the number and the profile of sources is unknown PCA can be employed to the large data sets[138]. It is based on the analysis of the correlation between measured

concentrations of chemical species in a number of samples, assuming that highly correlated compounds come from the same source[140]. However, such factor analysis model is limited due to the fact that it can generate negative profiles of species in a source as well as it does not account for the uncertainties in measurements. In addition, PCA can give qualitative information about the nature of the source profiles, but it does not provide a quantitative apportionment[138]. To solve all the limitations by the aforementioned receptor models, a useful tool in source apportionment called PMF was developed by EPA in 1994. This multivariate factor analysis model accounts for all the drawbacks of the previously mentioned models. Therefore, PMF has now become the most widely used source resolution model with more than 1000 papers stating its application[138, 141-144].

### **B-Principle of PMF: theoretical overview**

PMF Receptor model is based on the Chemical Mass Balance (CMB) equation that considers a single sample taken at a single location and time period, can be expressed as:

$$X_{ij} = \sum_{k=1}^p g_{ik} f_{kj} + e_{ij}$$

where,

$X_{ij}$  represents the concentration of ambient measured species  $j$  in sample  $i$

$P$  is the number of factors (sources) contributing to the measured sample and is provided to the model by the user

$f_{kj}$  is the fractional concentration of species  $j$  in the emissions from factor profile  $k$

$g_{ik}$  is the relative contribution of factor  $k$  to sample  $i$

$e_{ij}$  is the residual of the measured concentration of species  $j$  in sample  $i$  that cannot be explained by the model

In brief, the input matrix provided by the user is factorized into three matrices: factor contribution(G), factor profile(F), and residual (E).

The goal of solving this equation is to find values of  $g_{ik}$ ,  $f_{kj}$ , and  $p$  that best reproduce  $X_{ij}$ . This is accomplished by adjusting  $g_{ik}$ ,  $f_{kj}$  until a minimum value of  $Q_{robust}$  for a given  $p$  is found.  $Q_{robust}$  is the goodness of fit parameter calculated excluding outliers. However,  $Q_{true}$  is the goodness of fit parameter calculated including all collected data. For a stable and reliable solution  $Q_{robust}$  and  $Q_{true}$  should be more or less comparable to each other.  $Q_{robust}$  is expressed as:

$$Q_{robust} = \sum_{i=1}^n \sum_{j=1}^m \left[ \frac{x_{ij} - \sum_{k=1}^p g_{ik} f_{kj}}{u_{ij}} \right]^2$$

Thus the function  $Q_{robust}$  is basically the residues divided by uncertainties or is the weighted sum of squares of differences between the PMF output and the original data set. This means that in order to obtain a minimum  $Q$  that would give a feasible and robust solution, it is important that the residues which is the fraction that cannot be explained by the model to be low[144].

### **C-PMF model optimization and validation**

A critical step in PMF modeling is the determination of the correct number of factors/sources. In the optimization step, it is important to examine and respect five parameters in order to correctly estimate the number of factors. Thus, an optimal estimation

leads to a more stable, robust, and reliable solution that best reproduce the set of data originally provided by the user. The aforementioned five parameters are: signal to noise ratio, closeness of  $Q_{\text{robust}}$  to  $Q_{\text{true}}$ , residual analysis, correlation coefficients between observed and estimated concentrations and bootstrap mapping. In this study, a  $65 \times 16$  (65 samples with 16 PAHs each) was introduced into the EPA PMF 5.0. The number of factors from 3 to 6 was examined and the proximal number of factors was decided. According to the aforementioned parameters a 3-factor solution gave the lowest  $Q_{\text{robust}}$  1450 close to  $Q_{\text{true}}$  1459 ( $Q_{\text{robust}}/Q_{\text{true}} = 0.993$ ), a good scaled residuals range, best correlation coefficients and a good bootstrap mapping. The output results were easily interpretable and comparable to source profiles apportioned and reported in literature.

### **1-Signal to noise ratio (S/N):**

It indicates whether the variability in the measurements are real or within the noise of the data. If the data are within the noise, this indicates that our measurements are either near or below our detection limits. This is done by comparing our data to the calculated uncertainties which were provided by the user. The uncertainties can be calculated using multiple ways. One of the most common ways to calculate uncertainties is based on the limit of detection of the analytical method [145, 146] such that:

- When  $X_{ij} > \text{LOD}$ ,  $U_{ij} = 0.1X_{ij} + \text{LOD}/3$  Equation 1
- when  $X_{ij} < \text{LOD}$ ,  $U_{ij} = 0.2X_{ij} + \text{LOD}/3$  Equation 2

Because our measurements for the 16 PAHs were above the detection limit in all the samples Equation 1 was used for the calculation of the uncertainties. The strength of the species is examined by calculating their signal to the noise (S/N) ratio. Species are considered strong if

their S/N ratio is above 1, weak when S/N ratio is between 0.5 and 1, and bad when S/N ratio is below 0.5. According to our input data, all the species' strength were  $\geq 5.8$ , which are considered strong and thus, none of the 16 PAHs were excluded from the model.

## **2- Observed/ predicted scatter plots:**

Scatter plots compare observed (input data) values and predicted (modeled) values for each species to determine if the model fits the individual species well. In PMF, the minimum accepted correlation coefficient ( $R^2$ ) of one specie is 0.6[144]. for the 16 PAHs inputted to the model, the range of  $R^2$  was between 0.86 and 0.98 suggesting that the measured concentrations were well explained by three factor/source analysis.

## **3-Residual analysis**

This analysis indicates how much points were excluded from the solution of the model. If all residuals of the whole species are between +3 and -3 and they are normally distributed then this species is well modeled. If the residuals' range of a specie has a high scale and are non-normally distributed, it may be an indication of a poor fit. For the 16 PAHs, 87% of the scaled residuals estimated by PMF were normally distributed. Those which were beyond this range should be treated in caution while interpreting the output profiles[144].

## **4-Bootstrap mapping**

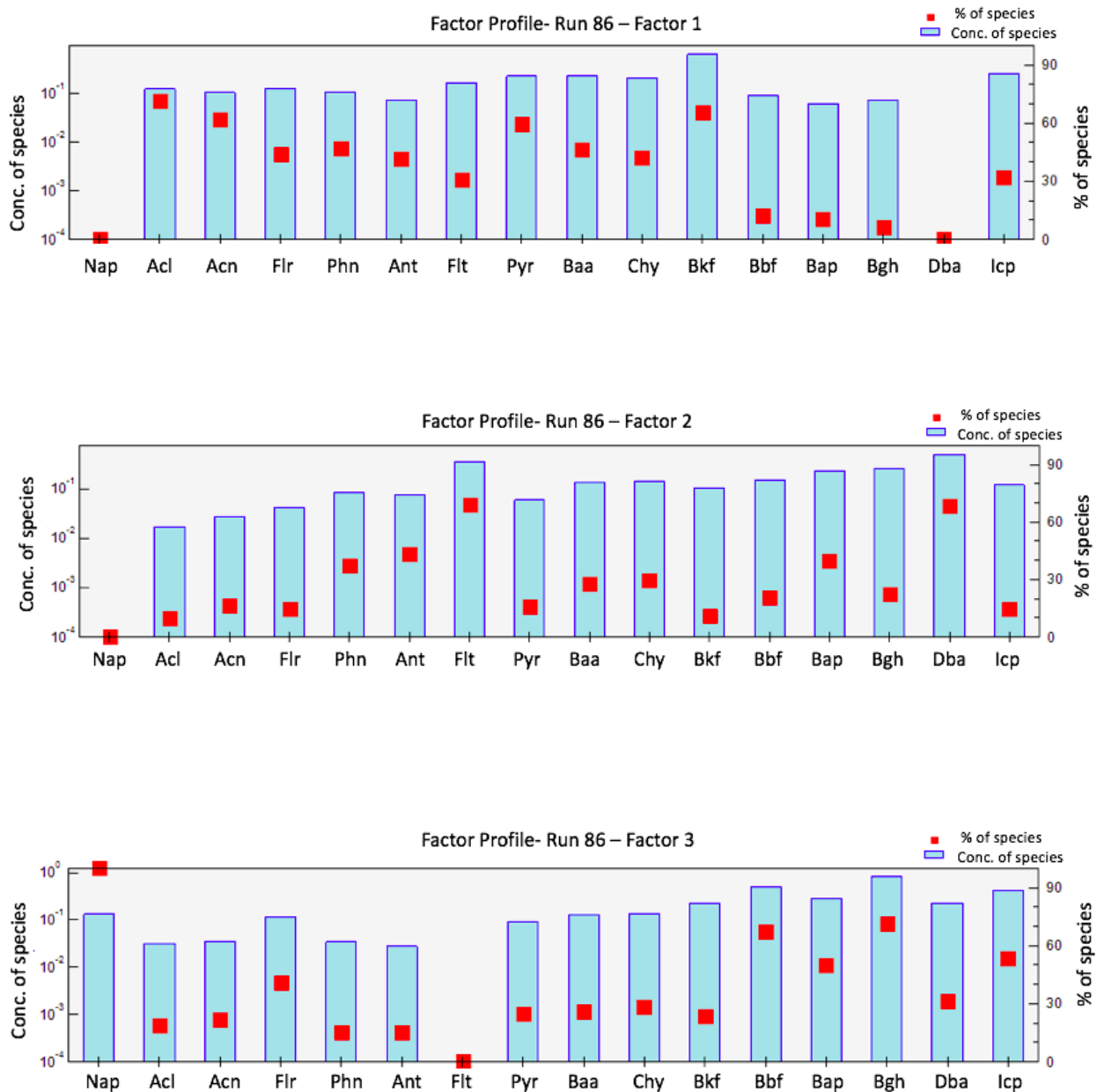
Is an indication whether the number of factors is approximate or has to be optimized[147-150]. A bootstrap model consists of running at least 100 bootstraps. The mapping percentage of each one of the assigned bootstrap factor to the base factor, indicates how unique are the different factor profiles. When a high number of a given bootstrapped factor highly overlap with more than one base factor, it may suggest that the factor profiles are not



unique and the number of factors may have to be optimized. In this study, the mapping percentage of the three base factors with the three bootstrap factors was over 80%, which indicates that the number of factor is appropriate.

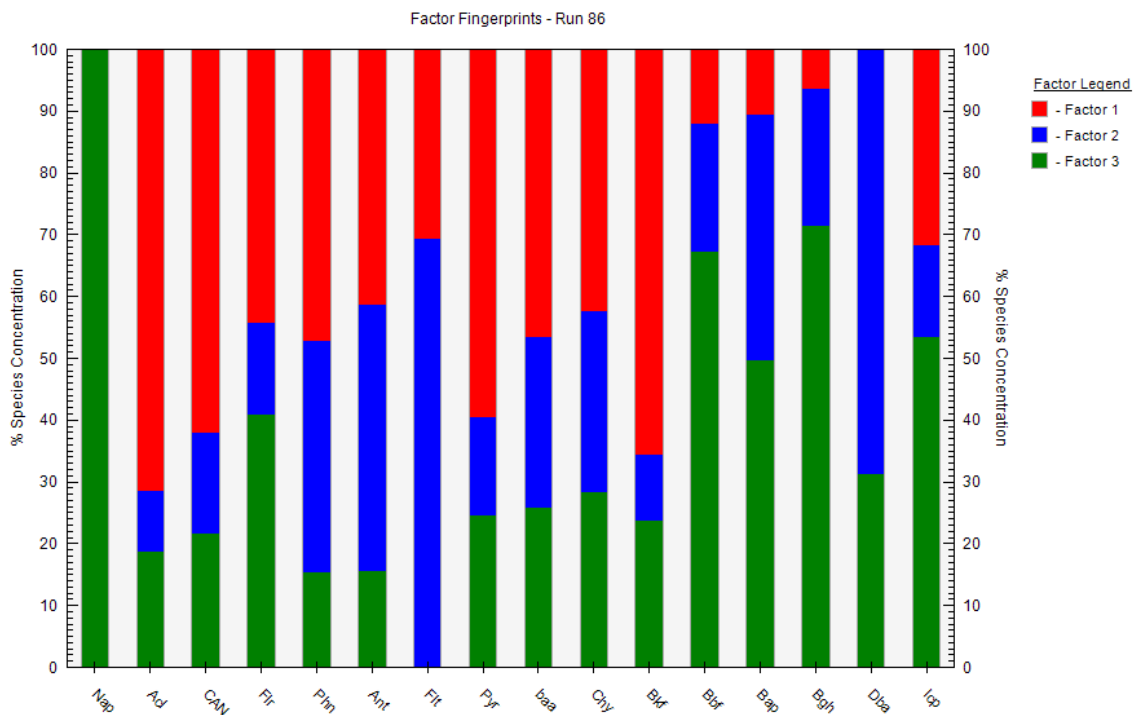
Once most of the PMF validation parameters were met, it is concluded that the appropriate number of factors affecting our study's receptor site is three. Thus, interpretation of the output follows.

## D-PMF output Results

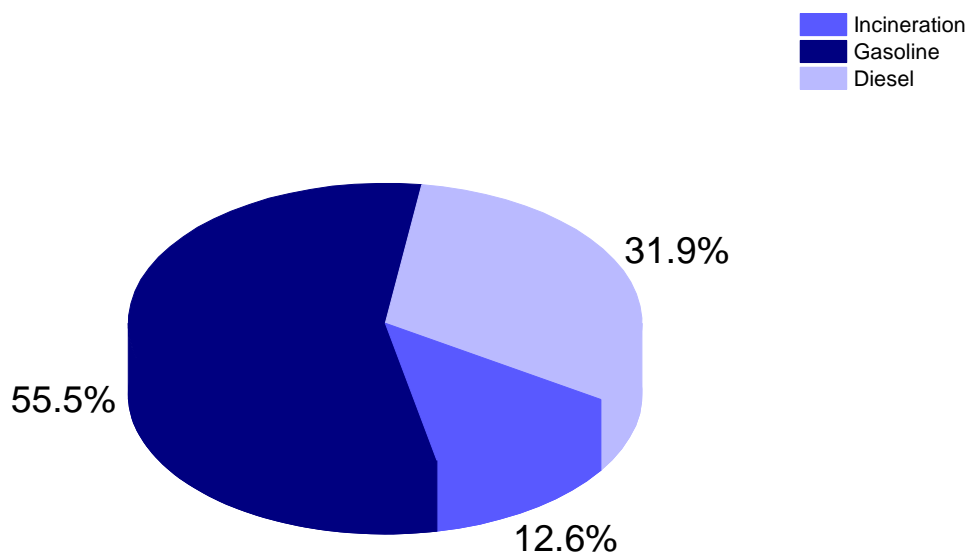


**Figure 30.** The three factor profiles for factors 1,2 and 3 determined by PMF and the fraction of the species in each. Nap(Naphthalene), Acl(Acenaphtylene), Acn(Acenaphtene), Flr(Flourene), Phn(Phenanthrene), Ant(Anthracene),

Flt(Flouranthene), Pyr(Pyrene), Baa( Benzo[a]anthracene), Chy(Chrysene),  
 Bkf(Benzo[k]flouranthene), Bbf(Benzo[b]flouranthene), Bap(Benzo[a]pyrene),  
 Bgh(Benzo[ghi]perylene), Dbh(Dibenzo[a,h]anthracene), Icp(Indeno[1,2,3-cd]pyrene)



**Figure 31.** Fingerprints (PAH markers) of the three factor profiles according to PMF results



**Figure 32.** The portion contribution of the three factors to the total ambient P-PAH

### **E-Results and Discussion:**

A three-factor model associated to P-PAHs gave the best fit in the PMF analysis. Mass contributions ( $\sum_{\text{total}} \text{PAH}$ ) were 31.9%, 12.6%, 55.5% for factors 1, 2 and 3 respectively.

Factor 1 (shown in Figure 30) accounted for 31.9% of the sum of the measured 16 PAHs. It has a high loading of 3-4 ring PAH characteristics which means that it is highly influenced by low molecular weight PAHs. The major PAHs contributing to factor 1 were pyrene 68%, acenaphthylene 70%, acenaphthene 80%, benzo[k]fluoranthene 70% and moderate contributions of flourene, phenanthrene, anthracene, chrysene, and benzo[a]anthracene. It is important to note that 5-6 member ring PAH contributions are negligible in this profile. It

has been revealed that diesel emissions are enriched in flouranthene, pyrene and benzo[k]flouranthene, chrysene relative to gasoline emissions[32, 39-42, 151].

Accordingly, factor one is attributed to diesel engine emissions. Diesel emissions are known to produce low molecular weight PAHs due to high temperature-induced cracking of hydrocarbons and formation of lower weight PAHs. In fact, in Beirut especially in hamra area, there is a continuous use of diesel generators as a quick fix to Lebanon's electricity problem. Electricite du Liban (EDL), the national electricity provider, has established a black-out rotating schedule of three hours per day for the area surrounding the sampling site. In fact, Shihadeh et al, have revealed that diesel generators contribute up to 40% of the total atmospheric P-PAH in Hamra area. The World Bank has reported that self-generation satisfies 33%–38% of electricity consumption in Lebanon[84]. In addition, the commercial ships that depart from and land in Beirut harbor have a serious impact on the total P-PAHs in the AUB area due to the high consumption of marine diesel oil by these ships.

Next, Factor two represents 12.6% of the sum of the measured 16 P-PAHs. This factor highly consists of dibenzo[ah]anthracene (72%) and flouranthene (71%) with moderate contribution of LMW P-PAHs (phenanthrene 35%, anthracene 50%) and HMW P-PAHs (Benzo(a) pyrene 40%) (see Figure 30). As reported by Zhao et al, Dibenzo[ah]anthracene and flouranthene are highly released by incinerators [152]. In fact, the medical waste incinerator of the American university hospital (AUH) located nearby AUB has an impact on the amount of factor two in the AUB site. is influenced by the medical waste incinerator at the AUH. Based on the information provided by the department of environment and health risk management (EHSRM) at AUB, about 3.5 % of cytotoxic medical waste

generated by the hospital is incinerated. The incinerator is known to operate for a maximum of six times per month with few hours at night[153].

Factor three contributes to 55.5% of all the measured 16 P-PAHs. High loadings of benzo[b]fluoranthene (70%), benzo[a]pyrene (52%), benzo[ghi]perylene (75%), indeno[cd]pyrene (57%) and naphthalene are observed. By visually comparing factor three to factor one (see Figure 30), we can conclude that factor three exhibits five to six-member ring P-PAHs, which means that factor three is influenced by HMW P-PAHs. Indeed, this kind of profile is usually attributed to gasoline emissions, which in AUB accounts for almost half of the P-PAHs[33, 34, 36-38]. The primary source of PAHs from gasoline is the on-road vehicles. In fact, AUB is surrounded 100 m West by the coastal Mediterranean road, a very busy street during rush hours, and 200 m East by Bliss street, a very busy street all day. Thus, no wonder why gasoline contributes to 55.5% of P-PAHs in the AUB area.

## Chapter V

# CONCLUSIONS

PM<sub>10</sub>-bound PAHs were measured during the year of 2017 covering all the seasons at an urban coastal site in Lebanon (AUB). The results show a high day to day variation even throughout the same season which were linked to the meteorological conditions that had a great impact on the dispersion or accumulation of P-PAHs. However, this variation between the samples decreased as we approached the spring and summer seasons, where both the weather and the emission sources are stagnant and stable in contrast to the cold period where the emissions are more ubiquitous due to seasonal emissions such as domestic heating and unstable weather. This variation was also clear in the individual P-PAHs, which showed a higher deviation during the cold period than the warm one. The HMW P-PAHs were the dominant P-PAHs at the site which reflect their low volatility rate and their origin from the most dominant source: gasoline emissions. Classification of P-PAHs showed that the 4-5-member ring P-PAHs were the only P-PAHs that had the highest contribution to the  $\Sigma$ P-PAH and exhibited a significant seasonal variation in contrast to the three and six-member ring, which reflect their seasonal combustion (pyrogenic) origin that is prominent in winter and absent in the other seasons. Source apportionment using PMF succeeded into evaluating the major sources of P-PAHs at the urban coastal site which were identified as gasoline, diesel and incineration with a portion contribution of 55.5%,31.2% and 12.6% respectively. The toxicity equivalent concentration of B[a]p was higher during the cold period.

This work was the first comprehensive study that succeeded into the determination of a P-PAH baseline at a representative site in Lebanon and into the evaluation of day to day and the seasonal variation of P-PAH at an urban coastal site exposed to several emission sources. In addition, this study represents a background dataset of P-PAHs that would be representative when comparing the levels to another urban location affected by an increase on the baseline level from specific emission sources. At the end, the cancer risk estimations showed that at ambient level exposure on a rooftop like AUB an adult individual would have a chance to develop cancer.



## BIBLIOGRAPHY:

1. Cancer, I.A.f.R.o., *IARC monographs on the evaluation of carcinogenic risk of chemicals to man*. IARC monographs on the evaluation of carcinogenic risk of chemicals to man., 1972. **1**.
2. Outlook, O.E., *OECD Environmental Outlook to 2050: The Consequences of Inaction*. 2012, Paris, France: OECD.
3. Organization, W.H., *Economic cost of the health impact of air pollution in Europe: Clean air, health and wealth*. World Health Organization, 2015. **2**.
4. EPA. *NAAQS Table*. 1990; Available from: <https://www.epa.gov/criteria-air-pollutants/naaqs-table>.
5. Organization, W.H., *Health effects of particulate matter. Policy implications for countries in Eastern Europe. Caucasus and central Asia*. World Health Organization Regional Office for Europe, Copenhagen, 2013.
6. Valavanidis, A., K. Fiotakis, and T. Vlachogianni, *Airborne particulate matter and human health: toxicological assessment and importance of size and composition of particles for oxidative damage and carcinogenic mechanisms*. *Journal of Environmental Science and Health, Part C*, 2008. **26**(4): p. 339-362.
7. Whitby, K., R. Husar, and B. Liu, *The aerosol size distribution of Los Angeles smog*. *Journal of Colloid and Interface Science*, 1972. **39**(1): p. 177-204.
8. Finlayson-Pitts, B.J. and J.N. Pitts Jr, *Chemistry of the upper and lower atmosphere: theory, experiments, and applications*. 1999: Academic press.
9. de Kok, T.M., et al., *Toxicological assessment of ambient and traffic-related particulate matter: a review of recent studies*. *Mutation Research/Reviews in Mutation Research*, 2006. **613**(2): p. 103-122.
10. Brown, J.S., et al., *Thoracic and respirable particle definitions for human health risk assessment*. *Particle and fibre toxicology*, 2013. **10**(1): p. 12.
11. Kim, K.-H., E. Kabir, and S. Kabir, *A review on the human health impact of airborne particulate matter*. *Environment International*, 2015. **74**(Supplement C): p. 136-143.
12. Löndahl, J., et al., *Size-resolved respiratory-tract deposition of fine and ultrafine hydrophobic and hygroscopic aerosol particles during rest and exercise*. *Inhalation toxicology*, 2007. **19**(2): p. 109-116.
13. Löndahl, J., et al. *Respiratory tract deposition of fine and ultrafine hydrophobic and hygroscopic aerosol particles during rest and exercise*. in *Nordic Society for Aerosol Research (NOSA) Annual Symposia, 2006*. 2006.
14. Fu, M., et al., *Advances of study on monitoring and evaluation of PM<sub>2.5</sub> pollution*. *Meteorology and Disaster Reduction Research*, 2011. **34**: p. 1-6.
15. Diociaiuti, M., et al., *The two PM<sub>2.5</sub> (fine) and PM<sub>2.5-10</sub> (coarse) fractions: evidence of different biological activity*. *Environmental research*, 2001. **86**(3): p. 254-262.

16. Jiménez, L.A., et al., *Activation of NF- $\kappa$ B by PM10 Occurs via an Iron-Mediated Mechanism in the Absence of I $\kappa$ B Degradation*. Toxicology and Applied Pharmacology, 2000. **166**(2): p. 101-110.
17. González-Flecha, B., *Oxidant mechanisms in response to ambient air particles*. Molecular Aspects of Medicine, 2004. **25**(1): p. 169-182.
18. Kadiiska, M.B., et al., *In vivo evidence of free radical formation in the rat lung after exposure to an emission source air pollution particle*. Chemical research in toxicology, 1997. **10**(10): p. 1104-1108.
19. Aust, A.E., et al., *Particle characteristics responsible for effects on human lung epithelial cells*. Research Report (Health Effects Institute), 2002(110): p. 1-65; discussion 67-76.
20. Hitzfeld, B., et al., *Airborne particulate matter modulates the production of reactive oxygen species in human polymorphonuclear granulocytes*. Toxicology, 1997. **120**(3): p. 185-195.
21. Heintzenberg, J., *Fine particles in the global troposphere A review*. Tellus B: Chemical and Physical Meteorology, 1989. **41**(2): p. 149-160.
22. Chow, J.C. and J.G. Watson, *Review of measurement methods and compositions for ultrafine particles*. Aerosol and Air Quality Research, 2007. **7**(2): p. 121-173.
23. Kelly, F.J. and J.C. Fussell, *Size, source and chemical composition as determinants of toxicity attributable to ambient particulate matter*. Atmospheric Environment, 2012. **60**(Supplement C): p. 504-526.
24. Chow, J.C. and J.G. Watson, *Survey of measurement and composition of ultrafine particles*. Aerosol and Air Quality Research, 2007. **7**(2): p. 121-173.
25. Organization, W.H., *WHO guidelines for indoor air quality: selected pollutants*. 2010.
26. Ross, J.A. and S. Nesnow, *Polycyclic aromatic hydrocarbons: correlations between DNA adducts and ras oncogene mutations*. Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis, 1999. **424**(1): p. 155-166.
27. Abdel-Shafy, H.I. and M.S.M. Mansour, *A review on polycyclic aromatic hydrocarbons: Source, environmental impact, effect on human health and remediation*. Egyptian Journal of Petroleum, 2016. **25**(1): p. 107-123.
28. IARC, *Some Non-heterocyclic Polycyclic Aromatic Hydrocarbons and Some Related Exposures*. 2010.
29. Ravindra, K., R. Sokhi, and R. Van Grieken, *Atmospheric polycyclic aromatic hydrocarbons: Source attribution, emission factors and regulation*. Atmospheric Environment, 2008. **42**(13): p. 2895-2921.
30. IARC. *AIR POLLUTION AND CANCER*. 2013; Available from: <https://www.iarc.fr/en/publications/books/sp161/AirPollutionandCancer161.pdf>.
31. Harrison, R.M., D. Smith, and L. Luhana, *Source apportionment of atmospheric polycyclic aromatic hydrocarbons collected from an urban location in Birmingham, UK*. Environmental Science & Technology, 1996. **30**(3): p. 825-832.
32. Ravindra, K., et al., *Seasonal and site-specific variation in vapour and aerosol phase PAHs over Flanders (Belgium) and their relation with anthropogenic activities*. Atmospheric Environment, 2006. **40**(4): p. 771-785.

33. Duval, M.M. and S.K. Friedlander, *Source resolution of polycyclic aromatic hydrocarbons in the Los Angeles atmosphere: application of a chemical species balance method with first order chemical decay. Final report Jan-Dec 80*. 1981, California Univ., Los Angeles (USA). Dept. of Chemical, Nuclear, and Thermal Engineering.
34. Masclat, P., G. Mouvier, and K. Nikolaou, *Relative decay index and sources of polycyclic aromatic hydrocarbons*. Atmospheric Environment (1967), 1986. **20**(3): p. 439-446.
35. Khalili, N.R., P.A. Scheff, and T.M. Holsen, *PAH source fingerprints for coke ovens, diesel and, gasoline engines, highway tunnels, and wood combustion emissions*. Atmospheric environment, 1995. **29**(4): p. 533-542.
36. Guo, H., et al., *Particle-associated polycyclic aromatic hydrocarbons in urban air of Hong Kong*. Atmospheric Environment, 2003. **37**(38): p. 5307-5317.
37. Miguel, A.H., et al., *On-road emissions of particulate polycyclic aromatic hydrocarbons and black carbon from gasoline and diesel vehicles*. Environmental Science & Technology, 1998. **32**(4): p. 450-455.
38. Marr, L.C., et al., *Characterization of polycyclic aromatic hydrocarbons in motor vehicle fuels and exhaust emissions*. Environmental science & technology, 1999. **33**(18): p. 3091-3099.
39. Caricchia, A.M., S. Chiavarini, and M. Pezza, *Polycyclic aromatic hydrocarbons in the urban atmospheric particulate matter in the city of Naples (Italy)*. Atmospheric Environment, 1999. **33**(23): p. 3731-3738.
40. Yang, H.-H., et al., *PAH emission from various industrial stacks*. Journal of Hazardous Materials, 1998. **60**(2): p. 159-174.
41. Ho, K., S. Lee, and G.M. Chiu, *Characterization of selected volatile organic compounds, polycyclic aromatic hydrocarbons and carbonyl compounds at a roadside monitoring station*. Atmospheric Environment, 2002. **36**(1): p. 57-65.
42. Omar, N.Y.M., et al., *Concentrations of PAHs in atmospheric particles (PM-10) and roadside soil particles collected in Kuala Lumpur, Malaysia*. Atmospheric Environment, 2002. **36**(2): p. 247-254.
43. Fang, G.-C., et al., *Polycyclic aromatic hydrocarbons study in Taichung, Taiwan, during 2002–2003*. Atmospheric Environment, 2004. **38**(21): p. 3385-3391.
44. Chmielewski, A.G., A. Ostapczuk, and J. Licki, *Electron beam technology for multipollutant emissions control from heavy fuel oil-fired boiler*. Journal of the Air & Waste Management Association, 2010. **60**(8): p. 932-938.
45. Yamasaki, H., K. Kuwata, and H. Miyamoto, *Effects of ambient temperature on aspects of airborne polycyclic aromatic hydrocarbons*. Environmental Science & Technology, 1982. **16**(4): p. 189-194.
46. Thibodeaux, L.J., et al., *The effect of moisture on volatile organic chemical gas-to-particle partitioning with atmospheric aerosols—competitive adsorption theory predictions*. Atmospheric Environment. Part A. General Topics, 1991. **25**(8): p. 1649-1656.
47. Arey, J. and R. Atkinson, *Photochemical reactions of PAHs in the atmosphere. PAHs: an ecotoxicological perspective*, 2003: p. 47-63.

48. Finlayson-Pitts, B.J. and J.N. Pitts Jr, *CHAPTER 10 - Airborne Polycyclic Aromatic Hydrocarbons and Their Derivatives: Atmospheric Chemistry and Toxicological Implications*, in *Chemistry of the Upper and Lower Atmosphere*. 2000, Academic Press: San Diego. p. 436-546.
49. Esteve, W., H. Budzinski, and E. Villenave, *Relative rate constants for the heterogeneous reactions of NO<sub>2</sub> and OH radicals with polycyclic aromatic hydrocarbons adsorbed on carbonaceous particles. Part 2: PAHs adsorbed on diesel particulate exhaust SRM 1650a*. *Atmospheric Environment*, 2006. **40**(2): p. 201-211.
50. Esteve, W., H. Budzinski, and E. Villenave, *Relative rate constants for the heterogeneous reactions of OH, NO<sub>2</sub> and NO radicals with polycyclic aromatic hydrocarbons adsorbed on carbonaceous particles. Part 1: PAHs adsorbed on 1–2 μm calibrated graphite particles*. *Atmospheric Environment*, 2004. **38**(35): p. 6063-6072.
51. Keyte, I.J., R.M. Harrison, and G. Lammel, *Chemical reactivity and long-range transport potential of polycyclic aromatic hydrocarbons—a review*. *Chemical Society Reviews*, 2013. **42**(24): p. 9333-9391.
52. Finlayson-Pitts, B.J. and J.N. Pitts Jr, *CHAPTER 1 - Overview of the Chemistry of Polluted and Remote Atmospheres*, in *Chemistry of the Upper and Lower Atmosphere*. 2000, Academic Press: San Diego. p. 1-14.
53. Sasaki, J., J. Arey, and W.P. Harger, *Formation of mutagens from the photooxidations of 2-4-ring PAH*. *Environmental science & technology*, 1995. **29**(5): p. 1324-1335.
54. Sasaki, J., et al., *Products of the gas-phase OH and NO<sub>3</sub> radical-initiated reactions of naphthalene*. *Environmental Science & Technology*, 1997. **31**(11): p. 3173-3179.
55. Lv, Y., et al., *Size distributions of polycyclic aromatic hydrocarbons in urban atmosphere: sorption mechanism and source contributions to respiratory deposition*. *Atmospheric Chemistry and Physics*, 2016. **16**(5): p. 2971-2983.
56. Akyüz, M. and H. Çabuk, *Meteorological variations of PM<sub>2.5</sub>/PM<sub>10</sub> concentrations and particle-associated polycyclic aromatic hydrocarbons in the atmospheric environment of Zonguldak, Turkey*. *Journal of Hazardous Materials*, 2009. **170**(1): p. 13-21.
57. Wu, S.P., S. Tao, and W.X. Liu, *Particle size distributions of polycyclic aromatic hydrocarbons in rural and urban atmosphere of Tianjin, China*. *Chemosphere*, 2006. **62**(3): p. 357-367.
58. Zelenyuk, A., et al., *Synergy between secondary organic aerosols and long-range transport of polycyclic aromatic hydrocarbons*. *Environmental science & technology*, 2012. **46**(22): p. 12459-12466.
59. Shrivastava, M., et al., *Global long-range transport and lung cancer risk from polycyclic aromatic hydrocarbons shielded by coatings of organic aerosol*. *Proceedings of the National Academy of Sciences*, 2017. **114**(6): p. 1246-1251.
60. Health, U.D.o. and H. Services, *Toxicological profile for polycyclic aromatic hydrocarbons*. Agency for Toxic Substances and Disease Registry, Atlanta, Ge., USA, 1995: p. 19.

61. IARC, *Some Non-heterocyclic Polycyclic Aromatic Hydrocarbons and Some Related Exposures* IARC Monographs on the Evaluation of Carcinogenic Risks to Humans , 2010. **92**.
62. Council, E.P.a., *arsenic, cadmium, mercury, nickel and polycyclic aromatic hydrocarbons in ambient air*. 2004.
63. Coleman, P., et al., *Assessment of benzo [a] pyrene atmospheric concentrations in the UK to support the establishment of a national PAH objective*. AEAT/ENV, 2001.
64. Organization, W.H., *Selected non-heterocyclic polycyclic aromatic hydrocarbons*. 1998.
65. Boström, C.-E., et al., *Cancer risk assessment, indicators, and guidelines for polycyclic aromatic hydrocarbons in the ambient air*. Environmental health perspectives, 2002. **110**(Suppl 3): p. 451.
66. Conney, A.H., *Induction of microsomal enzymes by foreign chemicals and carcinogenesis by polycyclic aromatic hydrocarbons: GHA Clowes Memorial Lecture*. Cancer research, 1982. **42**(12): p. 4875-4917.
67. Klingbeil, E., et al., *Polycyclic aromatic hydrocarbons, tobacco smoke, and epigenetic remodeling in asthma*. Immunologic research, 2014. **58**(2-3): p. 369-373.
68. Huang, H., et al., *Role of poly (ADP-ribose) glycohydrolase silencing in DNA hypomethylation induced by benzo (a) pyrene*. Biochemical and biophysical research communications, 2014. **452**(3): p. 708-714.
69. van Grevenynghe, J., et al., *Polycyclic aromatic hydrocarbons inhibit differentiation of human monocytes into macrophages*. The Journal of Immunology, 2003. **170**(5): p. 2374-2381.
70. Karakaya, A., I. Ates, and B. Yucesoy, *Effects of occupational polycyclic aromatic hydrocarbon exposure on T-lymphocyte functions and natural killer cell activity in asphalt and coke oven workers*. Human & experimental toxicology, 2004. **23**(7): p. 317-322.
71. Sanyal, M.K. and Y.L. Li, *Deleterious effects of polynuclear aromatic hydrocarbon on blood vascular system of the rat fetus*. Birth Defects Research Part B: Developmental and Reproductive Toxicology, 2007. **80**(5): p. 367-373.
72. Wormley, D., et al., *Inhaled benzo (a) pyrene impairs long-term potentiation in the F1 generation rat dentate gyrus*. Cellular and molecular biology (Noisy-le-Grand, France), 2004. **50**(6): p. 715-721.
73. Choi, H., et al., *Prenatal exposure to airborne polycyclic aromatic hydrocarbons and risk of intrauterine growth restriction*. Environmental health perspectives, 2008. **116**(5): p. 658.
74. Moorthy, B., C. Chu, and D.J. Carlin, *Polycyclic Aromatic Hydrocarbons: From Metabolism to Lung Cancer*. Toxicological Sciences, 2015. **145**(1): p. 5-15.
75. Saliba, N., et al., *Origin and variability of particulate matter (PM10 and PM2. 5) mass concentrations over an Eastern Mediterranean city*. Atmospheric Research, 2010. **97**(1-2): p. 106-114.
76. Massoud, R., et al., *Intraurban variability of PM10 and PM2. 5 in an Eastern Mediterranean city*. Atmospheric Research, 2011. **101**(4): p. 893-901.

77. Lelieveld, J., et al., *Global air pollution crossroads over the Mediterranean*. Science, 2002. **298**(5594): p. 794-799.
78. Saliba, N., et al., *Variation of selected air quality indicators over the city of Beirut, Lebanon: assessment of emission sources*. Atmospheric Environment, 2006. **40**(18): p. 3263-3268.
79. Saliba, N.A., H. Kouyoumdjian, and M. Roumié, *Effect of local and long-range transport emissions on the elemental composition of PM<sub>10-2.5</sub> and PM<sub>2.5</sub> in Beirut*. Atmospheric Environment, 2007. **41**(31): p. 6497-6509.
80. Saliba, N.A. and R. Massoud, *A comparative review of PM levels, sources, and their likely fates in the eastern Mediterranean region*, in *Urban Airborne Particulate Matter*. 2010, Springer. p. 3-17.
81. IPT, *Road Transport Sector and Air Pollution Case of Lebanon 2016*. 2016.
82. UN, *World Statistics Pocketbook 2017 edition*. 2017.
83. bank, w., *Supporting Lebanon's efforts to rebuild infrastructure and alleviate the impacts of conflict on municipalities*. 2013.
84. Bouri, E. and J. El Assad, *The Lebanese Electricity Woes: An Estimation of the Economical Costs of Power Interruptions*. Energies, 2016. **9**(8): p. 583.
85. LEBANON, I., *NATIONAL GREENHOUSE GAS INVENTORY REPORT AND MITIGATION ANALYSIS*. 2015.
86. Daher, N., et al., *Chemical composition of size-resolved particulate matter at near-freeway and urban background sites in the greater Beirut area*. Atmospheric Environment, 2013. **80**: p. 96-106.
87. Nakhlé, M.M., et al., *Short-term relationships between emergency hospital admissions for respiratory and cardiovascular diseases and fine particulate air pollution in Beirut, Lebanon*. Environmental monitoring and assessment, 2015. **187**(4): p. 196.
88. Daher, N., et al., *Oxidative potential and chemical speciation of size-resolved particulate matter (PM) at near-freeway and urban background sites in the greater Beirut area*. Science of the Total Environment, 2014. **470**: p. 417-426.
89. Shihadeh, A., et al., *Effect of distributed electric power generation on household exposure to airborne carcinogens in Beirut*. 2013.
90. Nassar, J., *SOURCES, DISPERSION AND TOXICITY OF PAHs AND DIOXINS AT AUB, ZOUK AND DORA*, in *Chemistry Department*. 2016, American University of Beirut: Beirut.
91. Teixeira, E.C., et al., *Polycyclic aromatic hydrocarbons study in atmospheric fine and coarse particles using diagnostic ratios and receptor model in urban/industrial region*. Environmental monitoring and assessment, 2013. **185**(11): p. 9587-9602.
92. Agudelo-Castañeda, D.M. and E.C. Teixeira, *Seasonal changes, identification and source apportionment of PAH in PM<sub>1.0</sub>*. Atmospheric environment, 2014. **96**: p. 186-200.
93. Akyüz, M. and H. Çabuk, *Meteorological variations of PM<sub>2.5</sub>/PM<sub>10</sub> concentrations and particle-associated polycyclic aromatic hydrocarbons in the atmospheric environment of Zonguldak, Turkey*. Journal of Hazardous Materials, 2009. **170**(1): p. 13-21.

94. Li, Z., et al., *Characterization of PM<sub>2.5</sub>-bound polycyclic aromatic hydrocarbons in Atlanta—seasonal variations at urban, suburban, and rural ambient air monitoring sites*. *Atmospheric Environment*, 2009. **43**(27): p. 4187-4193.
95. Li, Z., et al., *Characterization of PM<sub>2.5</sub>-bound polycyclic aromatic hydrocarbons in Atlanta*. *Atmospheric Environment*, 2009. **43**(5): p. 1043-1050.
96. Masiol, M., et al., *Carcinogenic and mutagenic risk associated to airborne particle-phase polycyclic aromatic hydrocarbons: a source apportionment*. *Atmospheric Environment*, 2012. **60**: p. 375-382.
97. Villar-Vidal, M., et al., *Air Polycyclic Aromatic Hydrocarbons (PAHs) associated with PM<sub>2.5</sub> in a North Cantabric coast urban environment*. *Chemosphere*, 2014. **99**: p. 233-238.
98. Lee, R.G., et al., *Emission factors and importance of PCDD/Fs, PCBs, PCNs, PAHs and PM<sub>10</sub> from the domestic burning of coal and wood in the UK*. *Environmental science & technology*, 2005. **39**(6): p. 1436-1447.
99. Deng, Y., et al., *The winter effect on formation of PCDD/Fs in Guangzhou by vehicles: A tunnel study*. *Atmospheric Environment*, 2011. **45**(15): p. 2541-2548.
100. Devos, O., et al., *EXHAUST EMISSIONS OF PAHs OF PASSENGER CARS*. *Polycyclic Aromatic Compounds*, 2006. **26**(1): p. 69-78.
101. Paturel, L., et al., *Analysis of PAH emissions from passenger cars by High Resolution Shpol'skii spectrofluorometry*. *Polycyclic Aromatic Compounds*, 1996. **9**(1-4): p. 331-339.
102. Tham, Y.W.F., K. Takeda, and H. Sakugawa, *Polycyclic aromatic hydrocarbons (PAHs) associated with atmospheric particles in Higashi Hiroshima, Japan: Influence of meteorological conditions and seasonal variations*. *Atmospheric Research*, 2008. **88**(3): p. 224-233.
103. Sharma, H., V. Jain, and Z.H. Khan, *Characterization and source identification of polycyclic aromatic hydrocarbons (PAHs) in the urban environment of Delhi*. *Chemosphere*, 2007. **66**(2): p. 302-310.
104. Olcese, L.E. and B.M. Toselli, *Some aspects of air pollution in Córdoba, Argentina*. *Atmospheric Environment*, 2002. **36**(2): p. 299-306.
105. Amarillo, A.C. and H. Carreras, *Quantifying the influence of meteorological variables on particle-bound PAHs in urban environments*. *Atmospheric Pollution Research*, 2016. **7**(4): p. 597-602.
106. Bucheli, T.D., et al., *Polycyclic aromatic hydrocarbons, black carbon, and molecular markers in soils of Switzerland*. *Chemosphere*, 2004. **56**(11): p. 1061-1076.
107. Wang, G., et al., *Identification, abundance and seasonal variation of anthropogenic organic aerosols from a mega-city in China*. *Atmospheric Environment*, 2007. **41**(2): p. 407-416.
108. Tham, Y.W., K. Takeda, and H. Sakugawa, *Polycyclic aromatic hydrocarbons (PAHs) associated with atmospheric particles in Higashi Hiroshima, Japan: Influence of meteorological conditions and seasonal variations*. *Atmospheric Research*, 2008. **88**(3-4): p. 224-233.

109. Valerio, F. and M. Pala, *Effects of temperature on the concentration of polycyclic aromatic hydrocarbons (PAHs) adsorbed onto airborne particulates*. Fresenius' journal of analytical chemistry, 1991. **339**(10): p. 777-779.
110. Zheng, M. and M. Fang, *Particle-associated polycyclic aromatic hydrocarbons in the atmosphere of Hong Kong*. Water, Air, and Soil Pollution, 2000. **117**(1-4): p. 175-189.
111. Duan, J., et al., *Seasonal variation on size distribution and concentration of PAHs in Guangzhou city, China*. Chemosphere, 2007. **67**(3): p. 614-622.
112. Venkataraman, C., S. Thomas, and P. Kulkarni, *Size distributions of polycyclic aromatic hydrocarbons—gas/particle partitioning to urban aerosols*. Journal of Aerosol Science, 1999. **30**(6): p. 759-770.
113. Hong, H., et al., *Seasonal variation of PM10-bound PAHs in the atmosphere of Xiamen, China*. Atmospheric Research, 2007. **85**(3-4): p. 429-441.
114. Rogge, W.F., et al., *Sources of fine organic aerosol. 2. Noncatalyst and catalyst-equipped automobiles and heavy-duty diesel trucks*. Environmental science & technology, 1993. **27**(4): p. 636-651.
115. Kavouras, I.G., et al., *Measurement of particulate aliphatic and polynuclear aromatic hydrocarbons in Santiago de Chile: source reconciliation and evaluation of sampling artifacts*. Atmospheric Environment, 1999. **33**(30): p. 4977-4986.
116. Mandalakis, M., et al., *Gas-particle concentrations and distribution of aliphatic hydrocarbons, PAHs, PCBs and PCDD/Fs in the atmosphere of Athens (Greece)*. Atmospheric Environment, 2002. **36**(25): p. 4023-4035.
117. Bi, X., et al., *Extractable organic matter in PM10 from LiWan district of Guangzhou City, PR China*. Science of the Total Environment, 2002. **300**(1-3): p. 213-228.
118. Manoli, E., A. Kouras, and C. Samara, *Profile analysis of ambient and source emitted particle-bound polycyclic aromatic hydrocarbons from three sites in northern Greece*. Chemosphere, 2004. **56**(9): p. 867-878.
119. Bourotte, C., et al., *A wintertime study of PAHs in fine and coarse aerosols in São Paulo city, Brazil*. Atmospheric Environment, 2005. **39**(21): p. 3799-3811.
120. del Rosario Sienra, M., N.G. Rosazza, and M. Préndez, *Polycyclic aromatic hydrocarbons and their molecular diagnostic ratios in urban atmospheric respirable particulate matter*. Atmospheric research, 2005. **75**(4): p. 267-281.
121. Wang, G., et al., *Aliphatic and polycyclic aromatic hydrocarbons of atmospheric aerosols in five locations of Nanjing urban area, China*. Atmospheric Research, 2006. **81**(1): p. 54-66.
122. Hong, H., et al., *Seasonal variation of PM10-bound PAHs in the atmosphere of Xiamen, China*. Atmospheric Research, 2007. **85**(3): p. 429-441.
123. Gogou, A., I. Bouloubassi, and E.G. Stephanou, *Marine organic geochemistry of the Eastern Mediterranean: 1. Aliphatic and polyaromatic hydrocarbons in Cretan Sea surficial sediments*. Marine Chemistry, 2000. **68**(4): p. 265-282.
124. Hwang, H.-M., T.L. Wade, and J.L. Sericano, *Concentrations and source characterization of polycyclic aromatic hydrocarbons in pine needles from Korea, Mexico, and United States*. Atmospheric Environment, 2003. **37**(16): p. 2259-2267.



125. Mostafa, A., et al., *Source characterization and the environmental impact of urban street dusts from Egypt based on hydrocarbon distributions*. Fuel, 2009. **88**(1): p. 95-104.
126. Takada, H., T. Onda, and N. Ogura, *Determination of polycyclic aromatic hydrocarbons in urban street dusts and their source materials by capillary gas chromatography*. Environmental Science & Technology, 1990. **24**(8): p. 1179-1186.
127. Ou, S., et al., *Petroleum hydrocarbons and polycyclic aromatic hydrocarbons in the surficial sediments of Xiamen Harbour and Yuan Dan Lake, China*. Chemosphere, 2004. **56**(2): p. 107-112.
128. Costa, H.J. and T.C. Sauer Jr, *Forensic approaches and considerations in identifying PAH background*. Environmental Forensics, 2005. **6**(1): p. 9-16.
129. Zakaria, M.P. and H. Takada, *Case study: oil spills in the strait of Malacca, Malaysia*. Oil Spill Environmental Forensics, Academic Press, Burlington, 2007: p. 489-504.
130. Stout, S., J. Leather, and W. Corl III, *A User's Guide for Determining the Sources of Contaminants in Sediments*. 2003, SPACE AND NAVAL WARFARE SYSTEMS CENTER SAN DIEGO CA.
131. Cancer, I.A.f.R.o., *Some non-heterocyclic polycyclic aromatic hydrocarbons and some related exposures*. Vol. 92. 2010: IARC Press, International Agency for Research on Cancer.
132. Pongpiachan, S., et al., *Assessing risks to adults and preschool children posed by PM<sub>2.5</sub>-bound polycyclic aromatic hydrocarbons (PAHs) during a biomass burning episode in Northern Thailand*. Science of the Total Environment, 2015. **508**: p. 435-444.
133. Pongpiachan, S., *Incremental lifetime cancer risk of PM<sub>2.5</sub> bound polycyclic aromatic hydrocarbons (PAHs) before and after the wildland fire episode*. Aerosol Air Qual. Res, 2016. **16**: p. 2907-2919.
134. Zhang, J., et al., *Estimating population exposure to ambient polycyclic aromatic hydrocarbon in the United States—Part II: Source apportionment and cancer risk assessment*. Environment international, 2016. **97**: p. 163-170.
135. Wang, J., et al., *Seasonal variation, spatial distribution and source apportionment for polycyclic aromatic hydrocarbons (PAHs) at nineteen communities in Xi'an, China: The effects of suburban scattered emissions in winter*. Environmental Pollution, 2017. **231**: p. 1330-1343.
136. Chen, S.-C. and C.-M. Liao, *Health risk assessment on human exposed to environmental polycyclic aromatic hydrocarbons pollution sources*. Science of the Total Environment, 2006. **366**(1): p. 112-123.
137. EPA, U., *Development of a Relative Potency Factor (RPF) Approach for Polycyclic Aromatic Hydrocarbon (PAH) Mixtures*. Intregrated Risk Information Systems (IRIS), 2010.
138. Hopke, P.K., *Review of receptor modeling methods for source apportionment*. Journal of the Air & Waste Management Association, 2016. **66**(3): p. 237-259.
139. Watson, J.G., et al., *The USEPA/DRI chemical mass balance receptor model, CMB 7.0*. Environmental Software, 1990. **5**(1): p. 38-49.

140. Wold, S., K. Esbensen, and P. Geladi, *Principal component analysis*. Chemometrics and intelligent laboratory systems, 1987. **2**(1-3): p. 37-52.
141. Hopke, P.K. *A guide to positive matrix factorization*. in *Workshop on UNMIX and PMF as Applied to PM2*. 2000.
142. Rose, K.A., *Source Allocation and Visibility Impairment in Two Class I Areas with Positive Matrix Factorization*. Report for US Environmental Protection Agency, Region, 2006. **10**.
143. Paatero, P. and U. Tapper, *Positive matrix factorization: A non- negative factor model with optimal utilization of error estimates of data values*. Environmetrics, 1994. **5**(2): p. 111-126.
144. Norris, G., et al., *EPA Positive Matrix Factorization (PMF) 5.0 fundamentals and User Guide Prepared for the US Environmental Protection Agency Office of Research and Development, Washington, DC*. 2014, DC EPA/600/R-14/108.
145. Jang, E., M.S. Alam, and R.M. Harrison, *Source apportionment of polycyclic aromatic hydrocarbons in urban air using positive matrix factorization and spatial distribution analysis*. Atmospheric environment, 2013. **79**: p. 271-285.
146. Tauler, R., et al., *Comparison of the results obtained by four receptor modelling methods in aerosol source apportionment studies*. Atmospheric Environment, 2009. **43**(26): p. 3989-3997.
147. Callén, M., et al., *Nature and sources of particle associated polycyclic aromatic hydrocarbons (PAH) in the atmospheric environment of an urban area*. Environmental Pollution, 2013. **183**: p. 166-174.
148. Gao, B., et al., *Source apportionment of atmospheric PAHs and their toxicity using PMF: impact of gas/particle partitioning*. Atmospheric environment, 2015. **103**: p. 114-120.
149. Callén, M.S., A. Iturmendi, and J.M. López, *Source apportionment of atmospheric PM2. 5-bound polycyclic aromatic hydrocarbons by a PMF receptor model. Assessment of potential risk for human health*. Environmental pollution, 2014. **195**: p. 167-177.
150. Sofowote, U.M., et al., *Assessing the long-range transport of PAH to a sub-Arctic site using positive matrix factorization and potential source contribution function*. Atmospheric environment, 2011. **45**(4): p. 967-976.
151. Ravindra, K., et al., *Assessment of air quality after the implementation of compressed natural gas (CNG) as fuel in public transport in Delhi, India*. Environmental monitoring and assessment, 2006. **115**(1-3): p. 405-417.
152. Zhao, L., et al., *Levels of polycyclic aromatic hydrocarbons in different types of hospital waste incinerator ashes*. Science of the total environment, 2008. **397**(1): p. 24-30.
153. Environmental health, S., and Risk management department. *AUH medical waste incineration*. 2017; Available from: <https://website.aub.edu.lb/facilities/ehsrn/Pages/index.aspx>.

